

Semantical and Algorithmic Aspects of the Living

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To tackle the complexity of biological systems, investigate:

- Programming Theory Concepts
- Formal Methods of Circuit and Program Verification
- Automated Reasoning Tools

Prototype implementation in the Biochemical Abstract Machine BIOCHAM v3.2
• modeling environment for Systems Biology

Systems Biology ?

“Systems Biology aims at systems-level understanding [of biological processes] which requires a set of principles and methodologies that links the behaviors of molecules to systems characteristics and functions.”

H. Kitano, ICSB 2000

Analyze post-genomic RNA and protein interaction data produced with high-throughput technologies available in databases like GO, KEGG, BioCyc, etc.

Integrate heterogeneous data about a specific problem

Understand and predict the behaviors of large networks of genes and proteins

- Systems Biology Markup Language (SBML): model exchange format
- Model repositories: e.g. biomodels.net 261 curated models of cell processes
- Simulation tools

Issue of Abstraction in Systems Biology

Models are built in Systems Biology with two contradictory perspectives :

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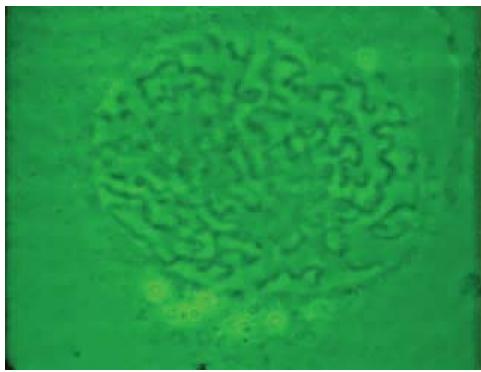
- 1) Models for representing knowledge : the more concrete the better
- 2) Models for making predictions : the more abstract the better !

These perspectives can be reconciled by organizing **models** and **formalisms** in **hierarchies of abstractions**.

To understand a system is not to know everything about it but to know abstraction levels that are sufficient for answering questions about it

Semantics of Living Processes ?

Formally, the semantics of a system depend on *our choice* of observables.



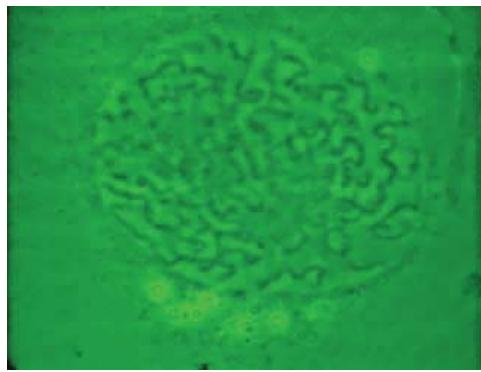
Mitosis movie [Lodish et al. 03]



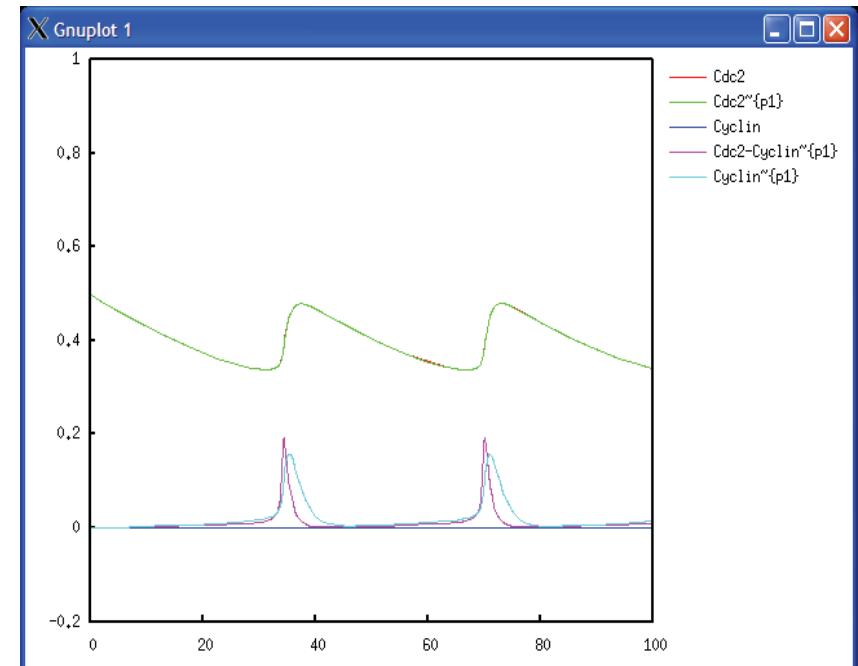
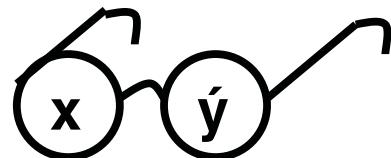
Continuous Differential Semantics

Formally, the semantics of a system depend on *our choice* of observables.

- Concentrations of molecules
- Reaction rates
- Ordinary Differential Equation (ODE) model



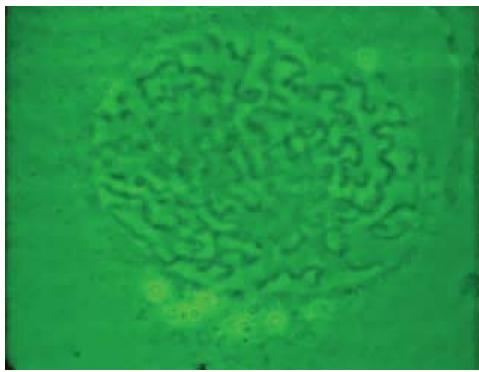
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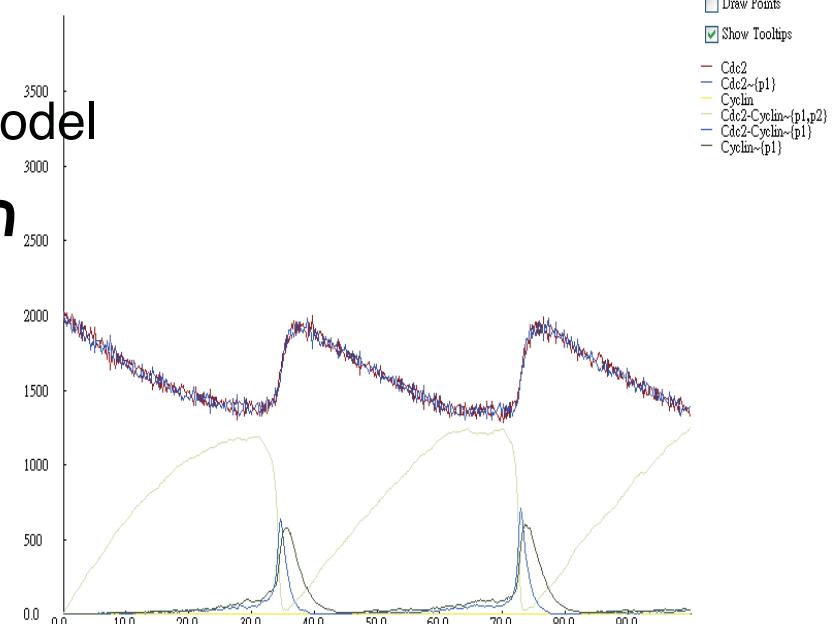
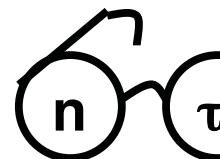
Stochastic Semantics

Formally, the semantics of a system depend on *our choice* of observables.

- (Small) numbers of molecules
- Probabilities of reaction
- Continuous Time Markov Chain (CTMC) model



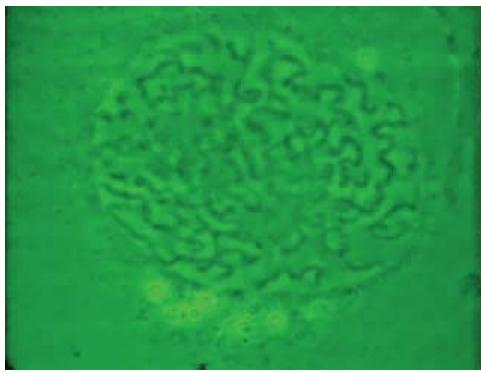
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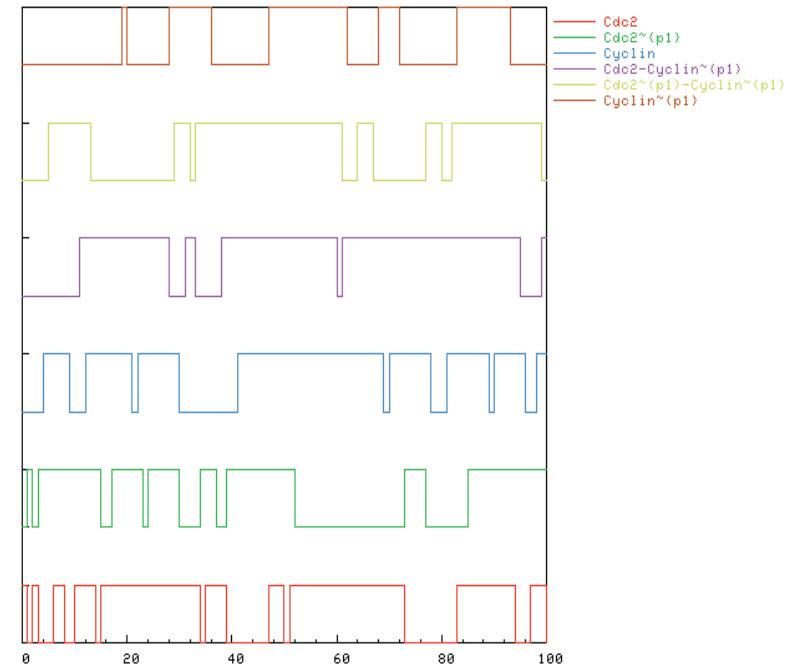
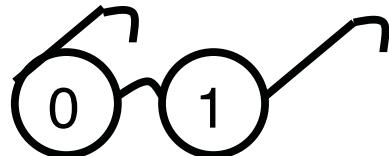
Boolean Semantics

Formally, the semantics of a system depend on *our choice* of observables.

- Presence/absence of molecules
- Boolean transition model



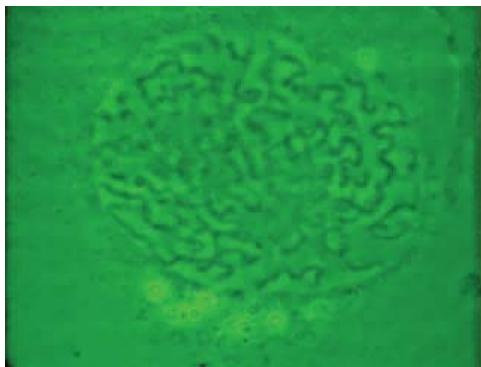
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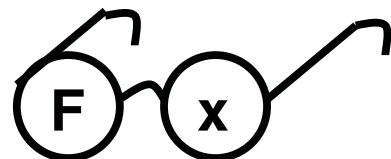
Propositional Temporal Logic

Formally, the semantics of a system depend on *our choice* of observables.

- Presence/absence of molecules
- Temporal logic formulas on Boolean traces



Mitosis movie [Lodish et al. 03]



F x

F (x \wedge F (\neg x \wedge y))

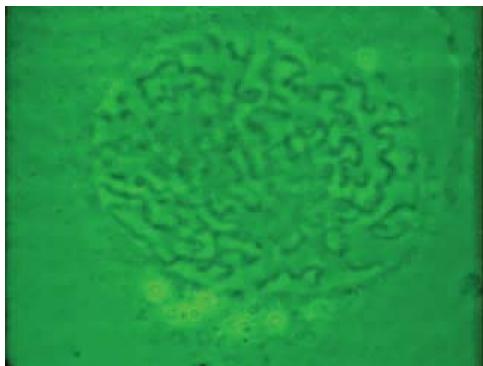
FG (x \vee y)

...

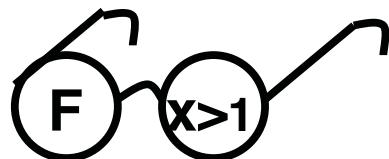
Constraint Temporal Logic LTL(R)

Formally, the semantics of a system depend on *our choice* of observables.

- Concentrations of molecules
- Temporal logic with constraints over R on quantitative traces



Mitosis movie [Lodish et al. 03]



$F(x > 0.2)$

$F(x > 0.2 \wedge F(x < 0.1 \wedge y > 0.2))$

$FG(x > 0.2 \vee y > 0.2)$

...

A Logical Paradigm for Systems Biology

Biological process model = (Quantitative) Transition System

Biological property = Temporal Logic Formula

Biological validation = Model-checking

Model inference = TL Constraint Solving

[Eker, Lincoln et al. PSB'02] [Chabrier Fages CMSB'03] [Bernot et al. TCS'04] ...

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Model:

- Boolean
- Differential
- Stochastic

BIOCHAM

- simulation
- query evaluation
- rule learning
- parameter learning
- robustness measure

↔

Biological Properties Specification:

- Temporal logic CTL
- LTL(R), QFLTL(R) constraints
- CSL

Overview of the Talk

1. Rule-based Modeling of Biochemical Systems

1. Syntax of molecules, compartments and reactions
2. Hierarchy of semantics: boolean, differential, stochastic
3. Cell cycle control models

2. *Temporal Logic Language* for formalizing Biological Properties

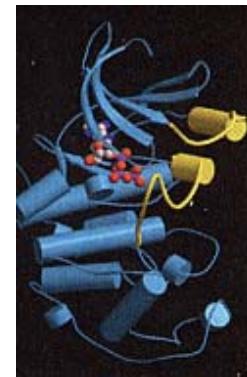
1. CTL for the boolean semantics
2. LTL(R) for the differential semantics
3. Parameter inference from LTL(R) properties
4. Robustness w.r.t. LTL(R) properties

3. *Conclusion* and collaborations

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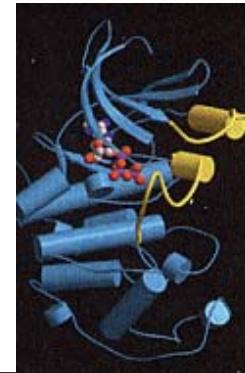
Formal Proteins: Syntax

- Cyclin dependent kinase 1 **Cdk1**
(free, inactive)



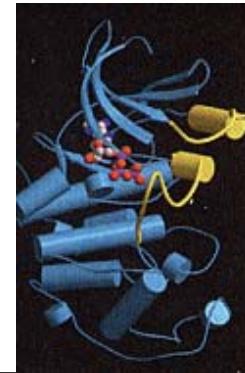
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- Complex Cdk1-Cyclin B **Cdk1–CycB**
(low activity)

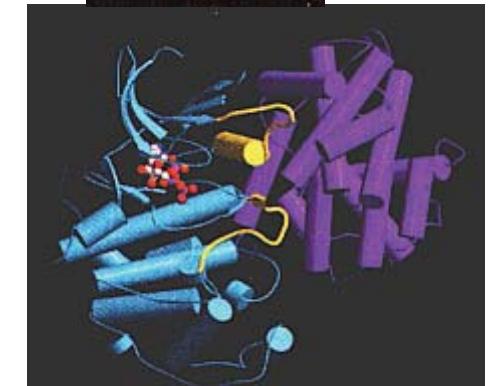


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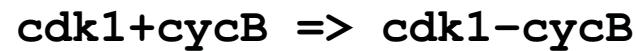


- Phosphorylated form $\text{Cdk1}^{\sim\{\text{thr161}\}}-\text{CycB}$
at site threonine 161
(high activity)
“Mitosis-Promoting Factor”
phosphorylates actin in microtubules → nuclear division



Formal Reaction Rules

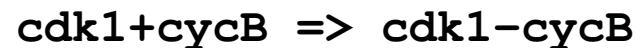
Complexation: $A + B \Rightarrow A-B.$



Decomplexation $A-B \Rightarrow A + B.$

Formal Reaction Rules

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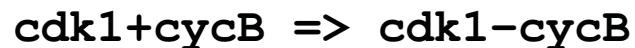
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Phosphorylation: $A = [K] \Rightarrow A\sim\{p\}.$



Formal Reaction Rules

Complexation: $A + B \Rightarrow A \cdot B$.

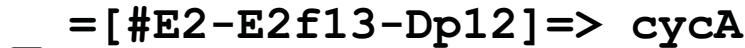


Decomplexation $A \cdot B \Rightarrow A + B$.

Phosphorylation: $A = [K] \Rightarrow A \sim \{p\}$.



Synthesis: $_ = [G] \Rightarrow A$.



Degradation: $A = [C] \Rightarrow _$.



(not for $cycE \cdot cdk2$ which is stable)

Formal Reaction Rules

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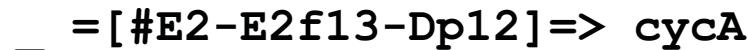


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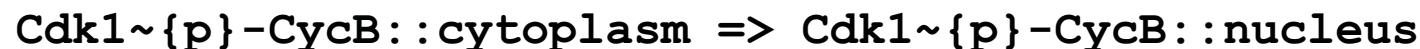


Degradation: $A = [C] \Rightarrow _$.



(not for $cycE \cdot cdk2$ which is stable)

Transport: $A::L1 \Rightarrow A::L2$.



Reaction Rule Models $\{ e_i \text{ for } l_i \Rightarrow r_i \}_{i \in I}$

SBML reaction rules with kinetics: $k * [A] * [B] \text{ for } A + B \Rightarrow C$

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Interpretations at four abstraction levels:

1. Differential Semantics: concentrations

- Ordinary Differential Equations
- Det. continuous hybrid automata

$$dx_k/dt = \sum_{i=1}^n r_i(x_k) * e_i - \sum_{j=1}^n l_j(x_k) * e_j$$

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2. Stochastic Semantics: numbers of molecules

- Continuous time Markov chain

$$p_{ij} = \frac{\tau_{ij}}{\sum_{(S_i, S_k, \tau_{ik}) \in s} \tau_{ik}}$$

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3. Discrete Semantics: numbers of molecules

- Multiset rewriting, Petri net
- CHAM [Berry Boudol 90] [Banatre Le Metayer 86]



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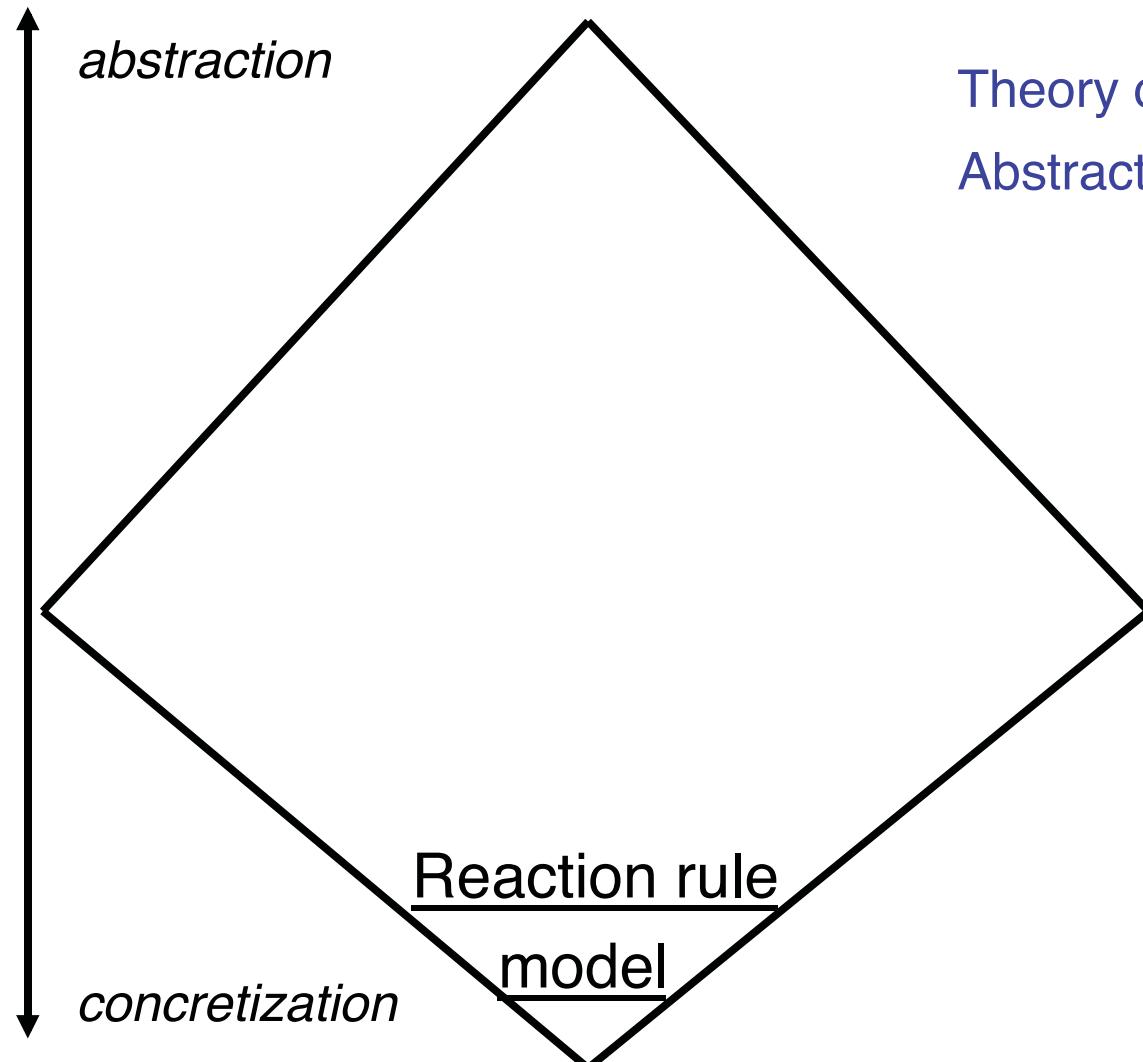


4. Boolean Semantics: presence-absence of molecules

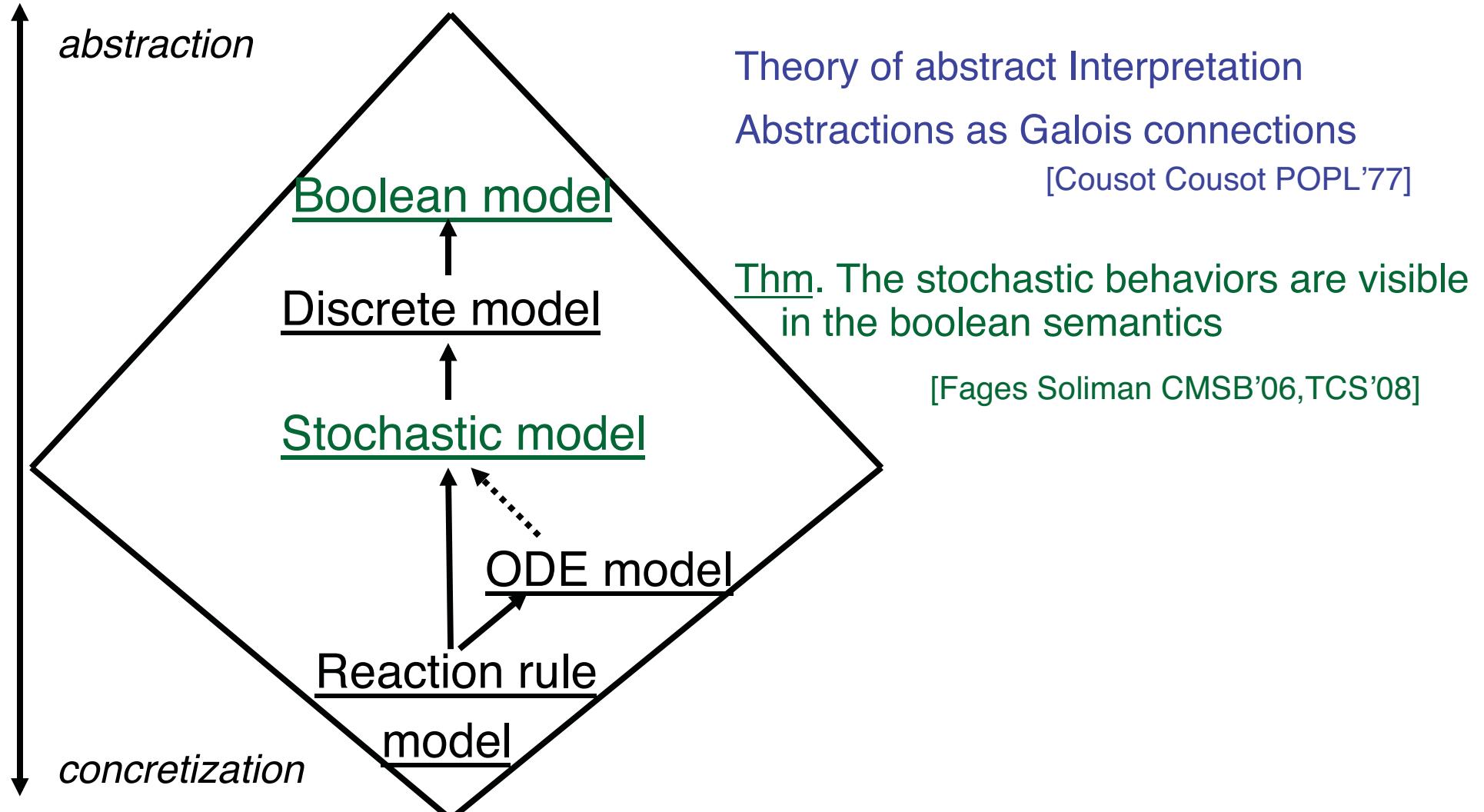
- Asynchronous Transition System



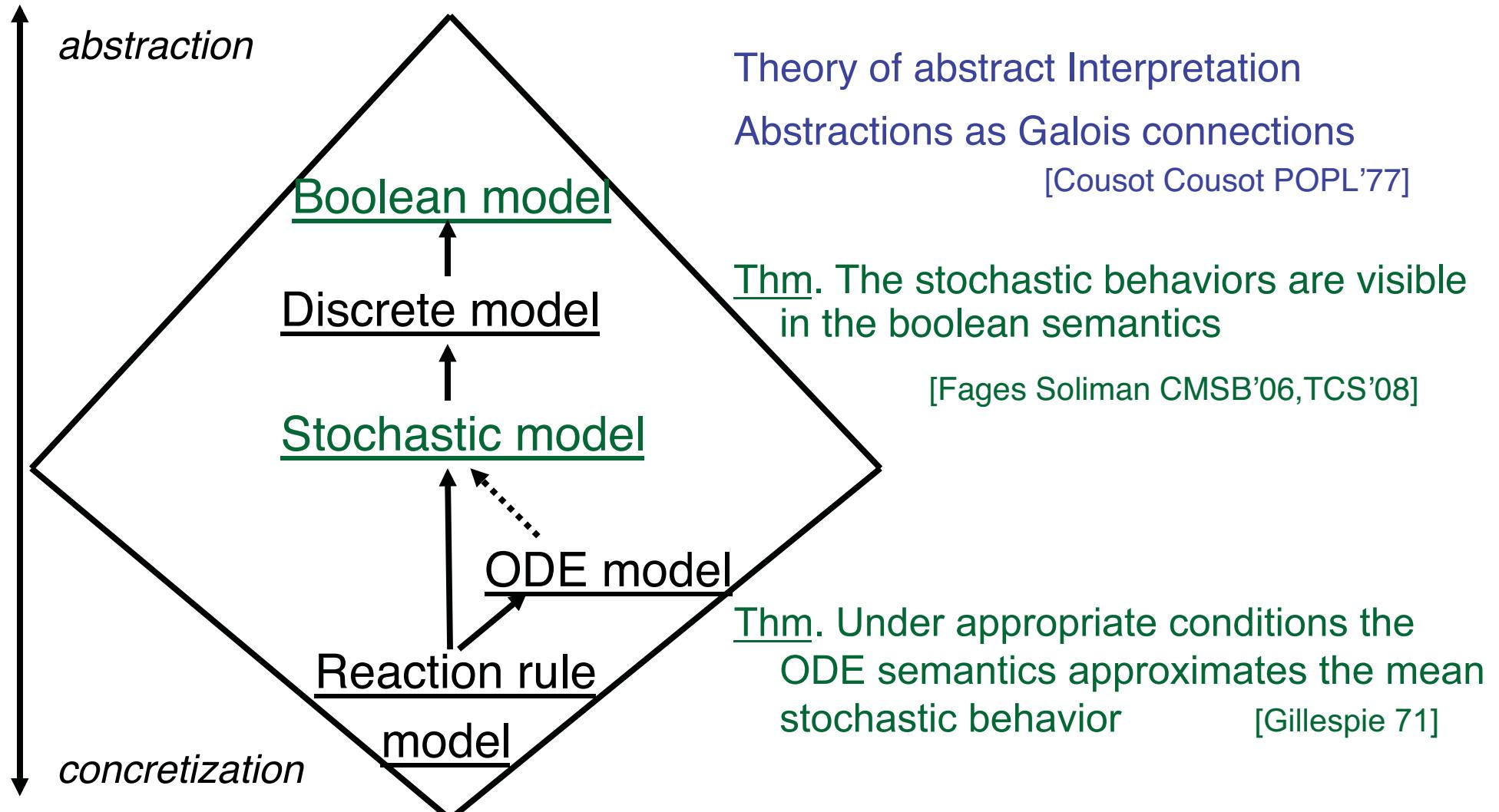
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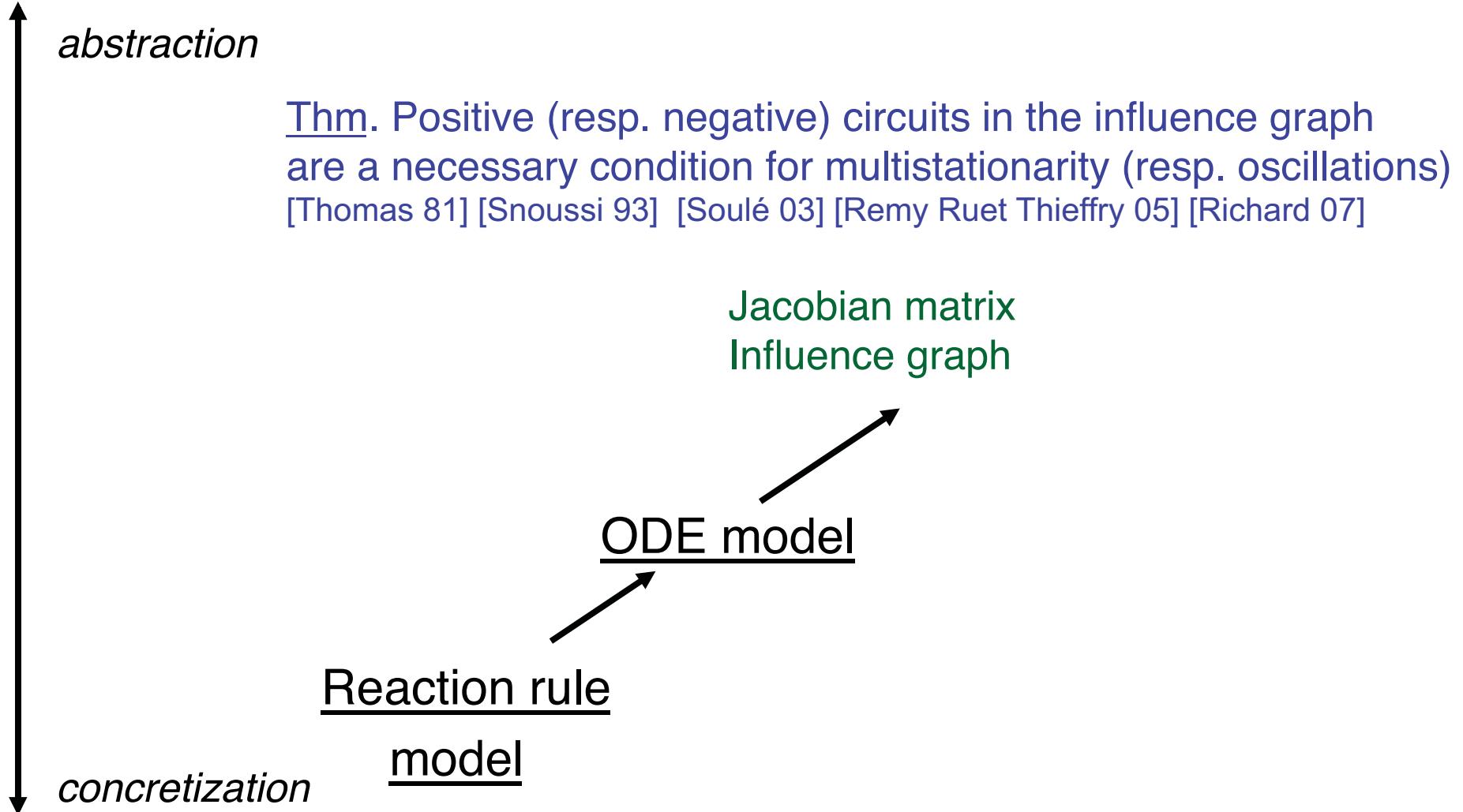
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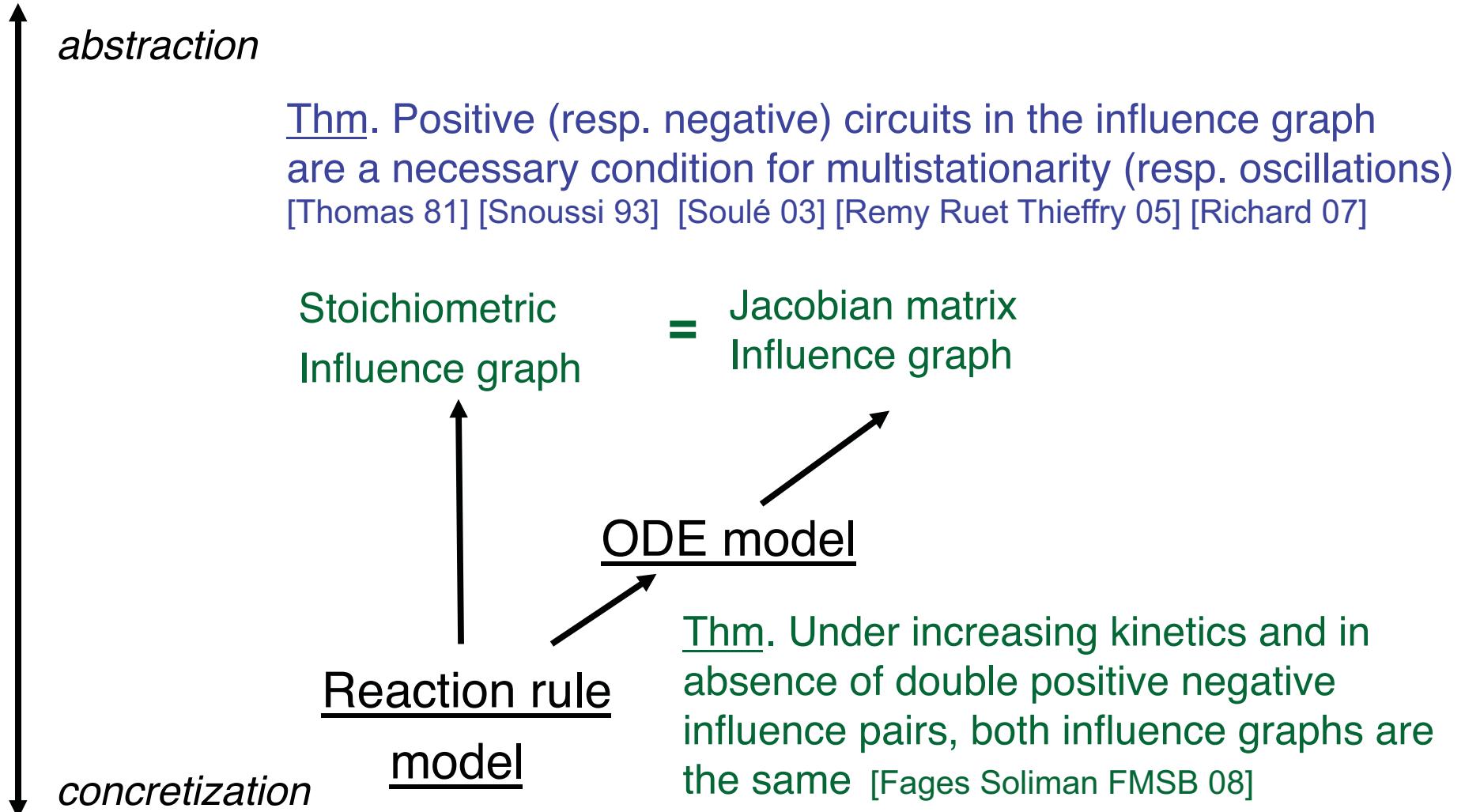
Hierarchy of Semantics



Influence Graph Abstraction



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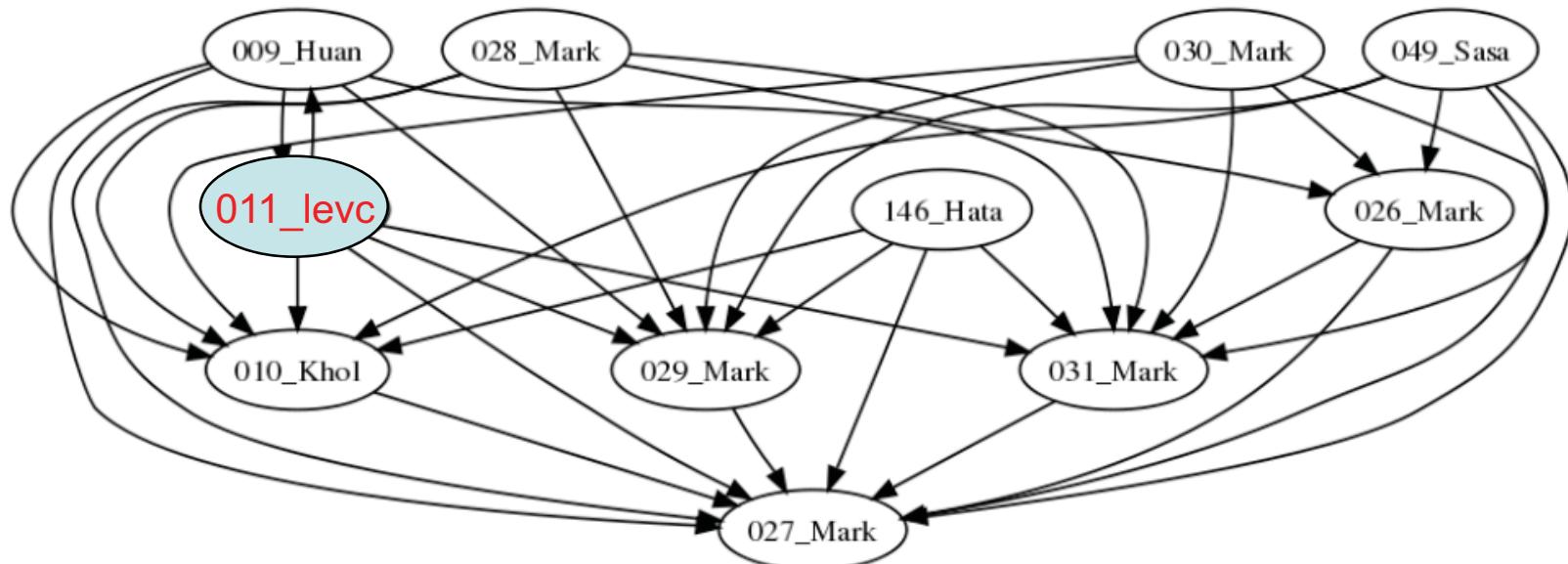


Hierarchies of Models

Model reductions as subgraph epimorphisms (constraint program)

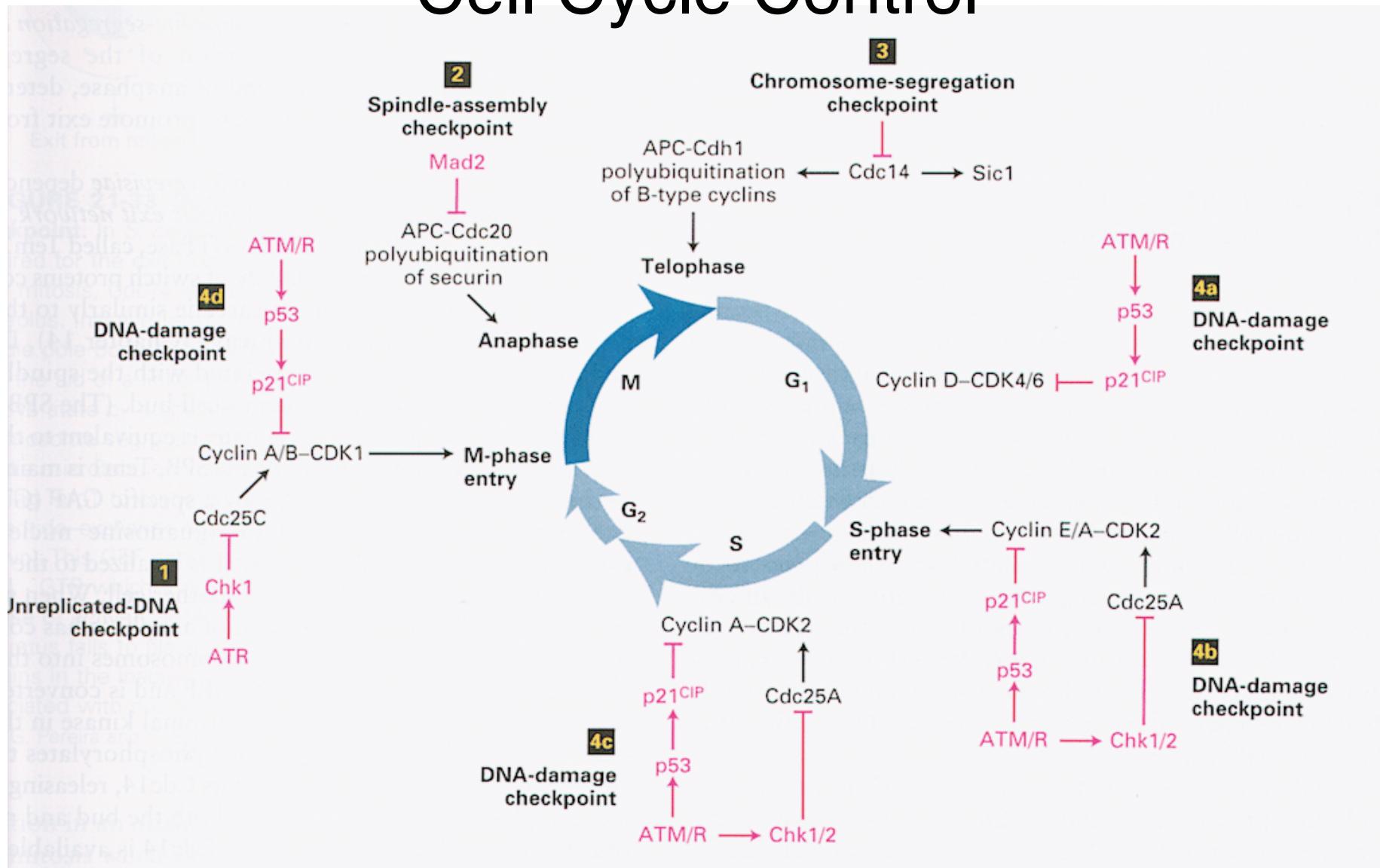
- 4 operations on reaction graphs: delete/merge species/reactions
- Query language for model repositories

[Gay Fages Soliman, A graph matching method for reducing and relating models 2010 bionfo]

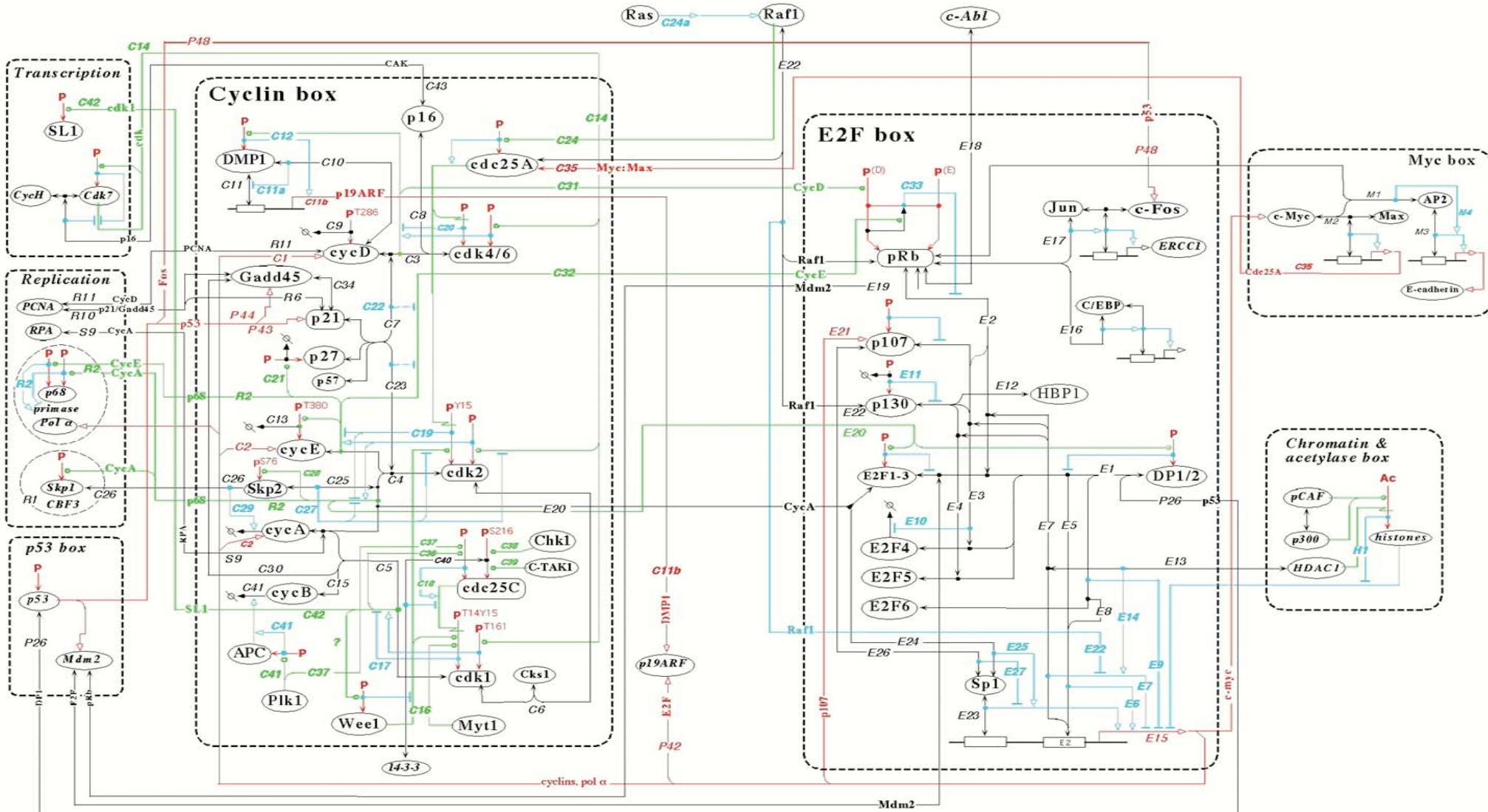


MAPK models from SBML model repository <http://www.biomodels.net>

Cell Cycle Control



Mammalian Cell Cycle Control Map [Kohn 99]



Kohn's map detail for Cdk2

Complexations with CycA and CycE

$\text{cdk2} \sim \$P + \text{cycA} - \$C \Rightarrow \text{cdk2} \sim \$P - \text{cycA} - \$C$

where \$C in {_, cks1} .

$\text{cdk2} \sim \$P + \text{cycE} \sim \$Q - \$C \Rightarrow \text{cdk2} \sim \$P - \text{cycE} \sim \$Q - \C

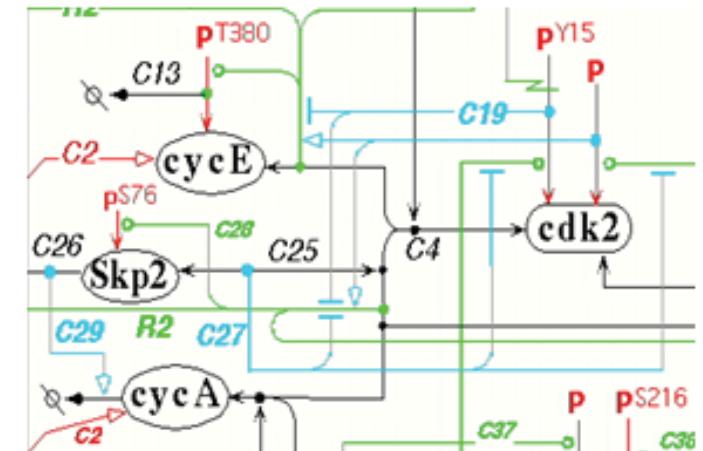
where \$C in {_, cks1} .

$p57 + \text{cdk2} \sim \$P - \text{cycA} - \$C \Rightarrow p57 - \text{cdk2} \sim \$P - \text{cycA} - \$C$

where \$C in {_, cks1} .

$\text{cycE} - \$C = [\text{cdk2} \sim \{p2\} - \text{cycE} - \$S] \Rightarrow \text{cycE} \sim \{\text{T380}\} - \C

where \$S in {_, cks1} and \$C in {_, cdk2 ~ ?, cdk2 ~ ? - cks1}



147 rule patterns \rightarrow 2733 expanded rules [Chabrier Chiaverini Danos Fages Schachter 04]

How to query the possible dynamical properties of such a system ?
 reachability, checkpoints, steady states, oscillations ?

Quantitative Model of Cell Cycle Control [Tyson 91]

k_1 for $_ \Rightarrow \text{Cyclin}$.

$k_2^*[\text{Cyclin}]$ for $\text{Cyclin} \Rightarrow _.$

$k_3^*[\text{Cyclin}]^*[\text{Cdc2}\sim\{\text{p1}\}]$ for
 $\text{Cyclin} + \text{Cdc2}\sim\{\text{p1}\} \Rightarrow \text{Cdc2}\sim\{\text{p1}\}-\text{Cyclin}\sim\{\text{p1}\}.$

$k_6^*[\text{Cdc2-Cyclin}\sim\{\text{p1}\}]$ for
 $\text{Cdc2-Cyclin}\sim\{\text{p1}\} \Rightarrow \text{Cdc2} + \text{Cyclin}\sim\{\text{p1}\}.$

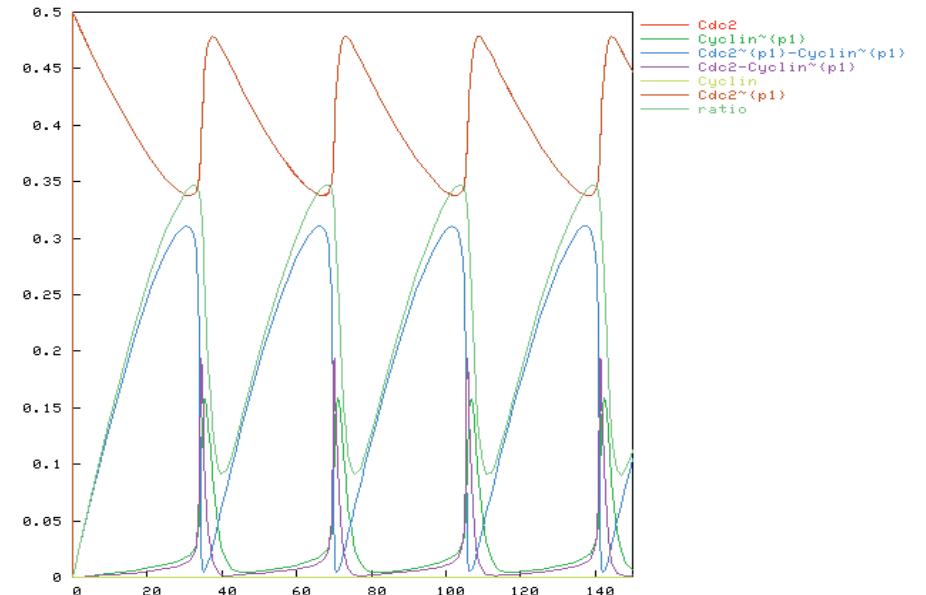
$k_7^*[\text{Cyclin}\sim\{\text{p1}\}]$ for $\text{Cyclin}\sim\{\text{p1}\} \Rightarrow _.$

$k_8^*[\text{Cdc2}]$ for $\text{Cdc2} \Rightarrow \text{Cdc2}\sim\{\text{p1}\}.$

$k_9^*[\text{Cdc2}\sim\{\text{p1}\}]$ for $\text{Cdc2}\sim\{\text{p1}\} \Rightarrow \text{Cdc2}.$

$k_{4p}^*[\text{Cdc2}\sim\{\text{p1}\}-\text{Cyclin}\sim\{\text{p1}\}]$ for
 $\text{Cdc2}\sim\{\text{p1}\}-\text{Cyclin}\sim\{\text{p1}\} \Rightarrow \text{Cdc2-Cyclin}\sim\{\text{p1}\}.$

$k_{4p}^*[\text{Cdc2-Cyclin}\sim\{\text{p1}\}]^2$ for
 $\text{Cdc2}\sim\{\text{p1}\}-\text{Cyclin}\sim\{\text{p1}\} = [\text{Cdc2-Cyclin}\sim\{\text{p1}\}] \Rightarrow \text{Cdc2-Cyclin}\sim\{\text{p1}\}.$



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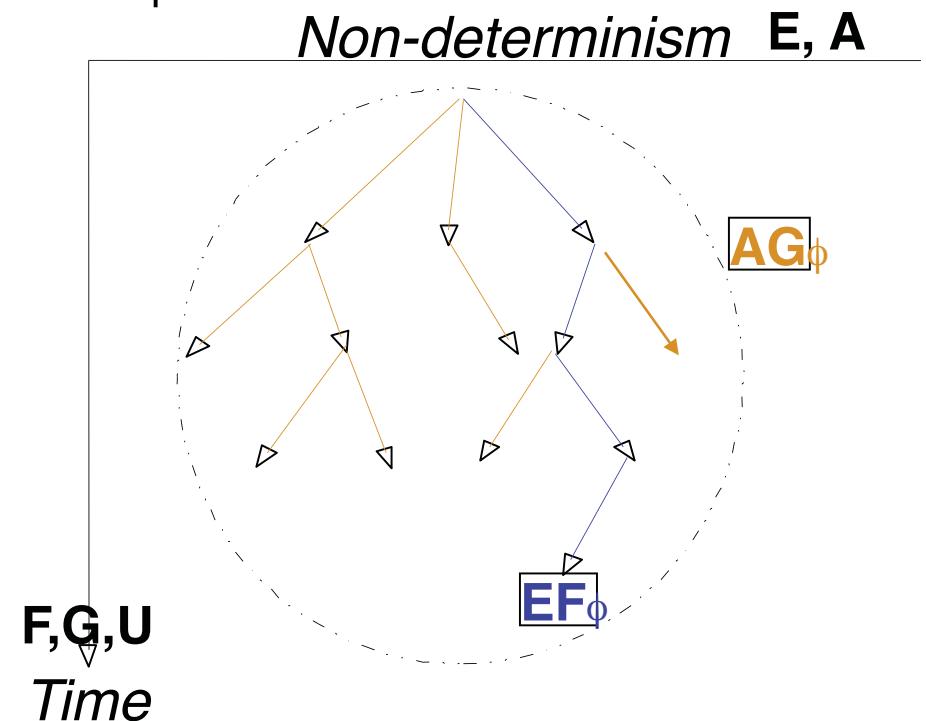
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Temporal Logic Queries

Temporal logics introduced for program verification by [Pnueli 77]

Computation Tree Logic CTL [Emerson Clarke 80]

Non-det. Time	E exists	A always
X next time	EX (φ)	AX (φ)
F finally	EF (φ) $\neg AG(\neg \varphi)$	AF (φ) <i>liveness</i>
G globally	EG (φ) $\neg AF(\neg \varphi)$	AG () <i>safety</i>
U until	E ($\varphi_1 \mathbf{U} \varphi_2$)	A ($\varphi_1 \mathbf{U} \varphi_2$)



Cell Cycle Model-Checking (with NuSMV Binary Decision Diagrams)

biocham: `check(reachable(cdk46~{p1,p2}-cycD~{p1})).`

Ei(EF(cdk46~{p1,p2}-cycD~{p1})) is true

biocham: `check(checkpoint(cdc25C~{p1,p2}, cdk1~{p1,p3}-cycB)).`

Ai(!(E(!(cdc25C~{p1,p2}) U cdk1~{p1,p3}-cycB))) is true

biocham: `nusmv(Ai(AG(!(cdk1~{p1,p2,p3}-cycB) -> checkpoint(Wee1, cdk1~{p1,p2,p3}-cycB))))).`

Ai(AG(!(cdk1~{p1,p2,p3}-cycB)->!(E(!(Wee1) U cdk1~{p1,p2,p3}-cycB)))) is false

biocham: `why.`

-- Loop starts here

cycB-cdk1~{p1,p2,p3} is present

cdk7 is present

cycH is present

cdk1 is present

Myt1 is present

cdc25C~{p1} is present

rule_114 cycB-cdk1~{p1,p2,p3}=[cdc25C~{p1}]=>cycB-cdk1~{p2,p3}.

cycB-cdk1~{p2,p3} is present

cycB-cdk1~{p1,p2,p3} is absent

rule_74 cycB-cdk1~{p2,p3}=[Myt1]=>cycB-cdk1~{p1,p2,p3}.

cycB-cdk1~{p2,p3} is absent

cycB-cdk1~{p1,p2,p3} is present

Kohn's Map Model-Checking

147-2733 rules, 165 proteins and genes, 500 variables, 2^{500} states.

Biocham NuSMV model-checker time in seconds [Chabrier Fages CMSB 03]

Initial state G2	Query:	Time:
	compiling	29
Reachability G1	EF CycE	2
Reachability G1	EF CycD	1.9
Reachability G1	EF PCNA-CycD	1.7
Checkpoint for mitosis complex	$\neg \mathbf{EF} (\neg \text{Cdc25}\sim\{\text{Nterm}\})$ $\mathbf{U} \text{Cdk1}\sim\{\text{Thr161}\}-\text{CycB}$	2.2
Oscillations CycA	EG ((EF \neg CycA) \wedge (EF CycA))	31.8
Oscillations CycB	EG ((EF \neg CycB) \wedge (EF CycB)) false !	6

Constraint Linear Time Logic LTL(R)

- $\mathbf{F}([A]>10)$: the concentration of A eventually gets above 10.
- $\mathbf{FG}([A]>10)$: the concentration of A eventually reaches and remains above value 10.
- $\mathbf{F}(\text{Time}=t_1 \wedge [A]=v_1 \wedge \mathbf{F}(\dots \wedge \mathbf{F}(\text{Time}=t_N \wedge [A]=v_N)\dots))$
Numerical data time series (e.g. experimental curves)
- $\mathbf{G}([A]+[B]<[C])$: the concentration of C is always greater than the sum of the concentrations of A and B.
- $\mathbf{F}([\text{M}] < V \wedge \mathbf{X}([\text{M}] = V) \wedge \mathbf{X}([\text{M}] < V)))$: local maximum V
- oscillations, period constraints, etc.

Quantitative Model of Cell Cycle Control [Tyson 91]

k_1 for $_ \Rightarrow \text{Cyclin}$.

$k_2 * [\text{Cyclin}] \text{ for } \text{Cyclin} \Rightarrow _.$

$k_3 * [\text{Cyclin}] * [\text{Cdc2} \sim \{p1\}] \text{ for }$
 $\text{Cyclin} + \text{Cdc2} \sim \{p1\} \Rightarrow \text{Cdc2} \sim \{p1\}-\text{Cyclin} \sim \{p1\}.$

$k_6 * [\text{Cdc2}-\text{Cyclin} \sim \{p1\}] \text{ for }$
 $\text{Cdc2}-\text{Cyclin} \sim \{p1\} \Rightarrow \text{Cdc2} + \text{Cyclin} \sim \{p1\}.$

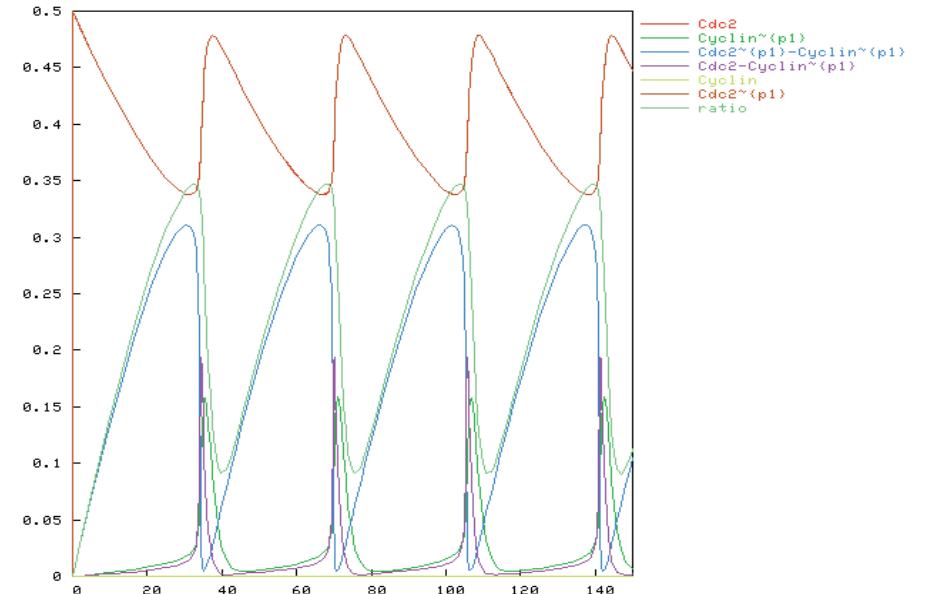
$k_7 * [\text{Cyclin} \sim \{p1\}] \text{ for } \text{Cyclin} \sim \{p1\} \Rightarrow _.$

$k_8 * [\text{Cdc2}] \text{ for } \text{Cdc2} \Rightarrow \text{Cdc2} \sim \{p1\}.$

$k_9 * [\text{Cdc2} \sim \{p1\}] \text{ for } \text{Cdc2} \sim \{p1\} \Rightarrow \text{Cdc2}.$

$k_{4p} * [\text{Cdc2} \sim \{p1\}-\text{Cyclin} \sim \{p1\}] \text{ for }$
 $\text{Cdc2} \sim \{p1\}-\text{Cyclin} \sim \{p1\} \Rightarrow \text{Cdc2}-\text{Cyclin} \sim \{p1\}.$

$k_{4*} [\text{Cdc2}-\text{Cyclin} \sim \{p1\}]^2 * [\text{Cdc2} \sim \{p1\}-\text{Cyclin} \sim \{p1\}] \text{ for }$
 $\text{Cdc2} \sim \{p1\}-\text{Cyclin} \sim \{p1\} = [\text{Cdc2}-\text{Cyclin} \sim \{p1\}] \Rightarrow \text{Cdc2}-\text{Cyclin} \sim \{p1\}.$



Numerical Integration of ODE Models

$$dX/dt = f(X).$$

Initial conditions X_0

Idea: discretize time $t_0, t_1=t_0+\Delta t_0, t_2=t_1+\Delta t_1, \dots$
and compute a numerical trace

$$(t_0, X_0), (t_1, X_1), \dots, (t_n, X_n) \dots$$

- *Euler's method:* $t_{i+1}=t_i + \Delta t$ $X_{i+1}=X_i+f(X_i)^*\Delta t$
error estimation $E(X_{i+1})=|f(X_i)-f(X_{i+1})|^*\Delta t$
- *Runge-Kutta's method:* intermediate computations at $\Delta t/2$
- *Adaptive step method:* $\Delta t_{i+1}=\Delta t_i/2$ while $E>E_{\max}$, otherwise $\Delta t_{i+1}=2^*\Delta t_i$
- *Rosenbrock's stiff method:* solve $X_{i+1}=X_i+f(X_{i+1})^*\Delta t$ by formal differentiation

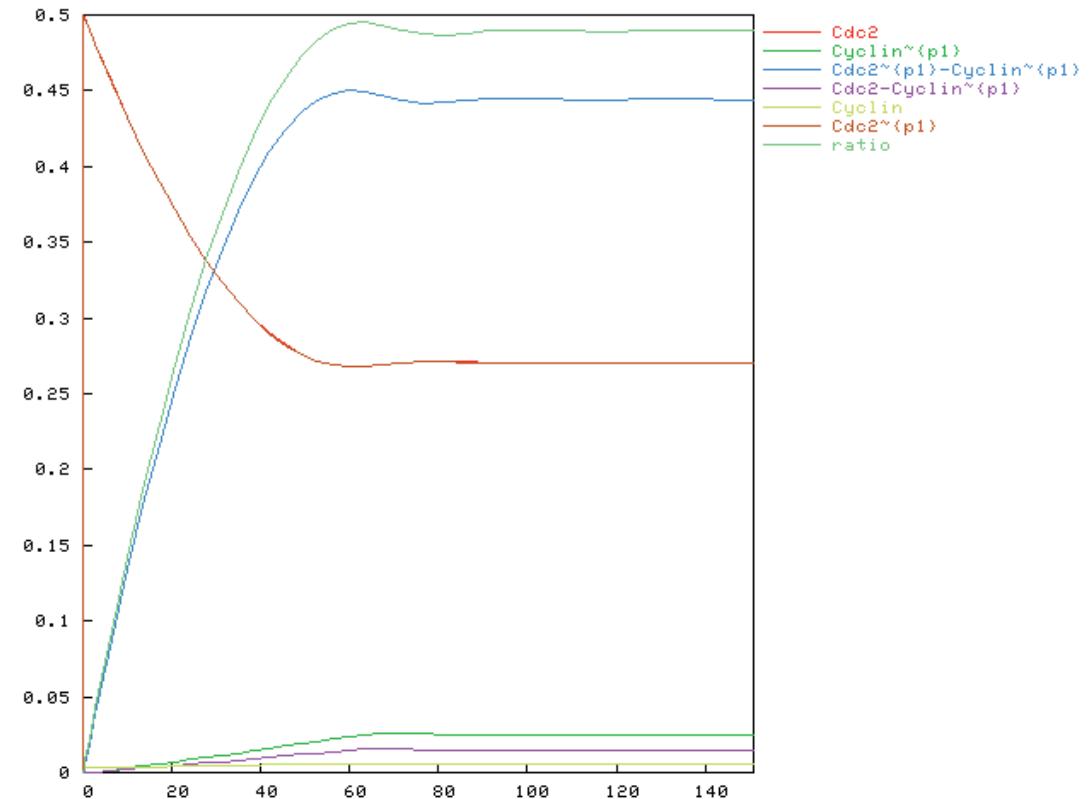
Parameter Search from Temporal Properties

```
biocham: learn_parameter([k3,k4],[(0,200),(0,200)],20,  
oscil(Cdc2-Cyclin~{p1},3),150).
```

Parameter Search from Temporal Properties

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biocham: learn_parameter([k3,k4],[(0,200),(0,200)],20,  
oscil(Cdc2-Cyclin~{p1}.3).150).
```

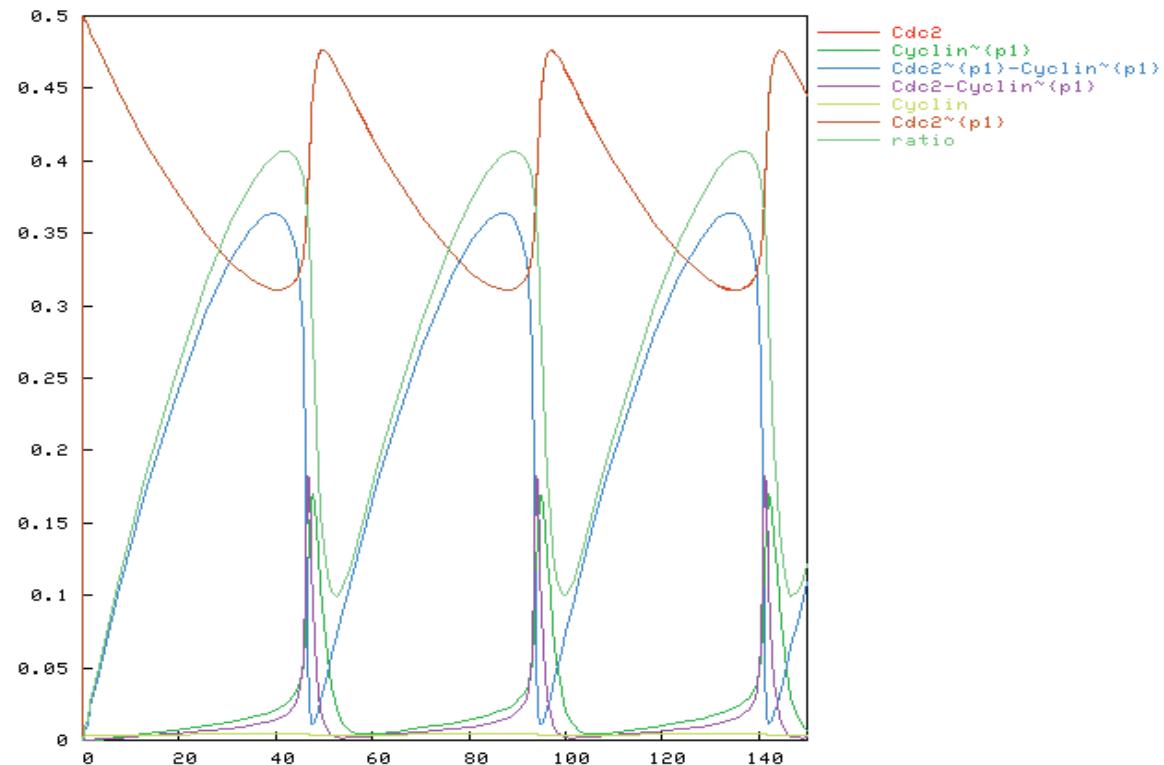
First values found :
parameter(k3,10).
parameter(k4,70).



Parameter Search from Temporal Properties

```
biocham: learn_parameter([k3,k4],[(0,200),(0,200)],20,  
oscil(Cdc2-Cyclin~{p1},3) & F([Cdc2-Cyclin~{p1}]>0.15), 150).
```

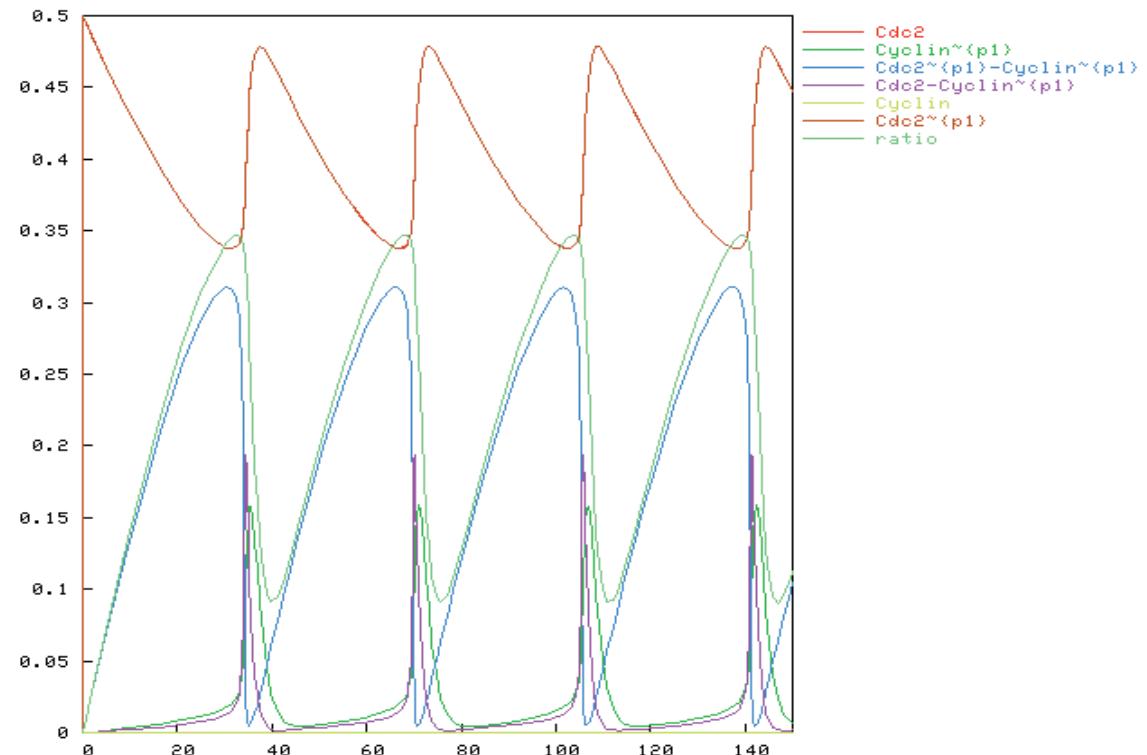
First values found :
parameter(k3,10).
parameter(k4,120).



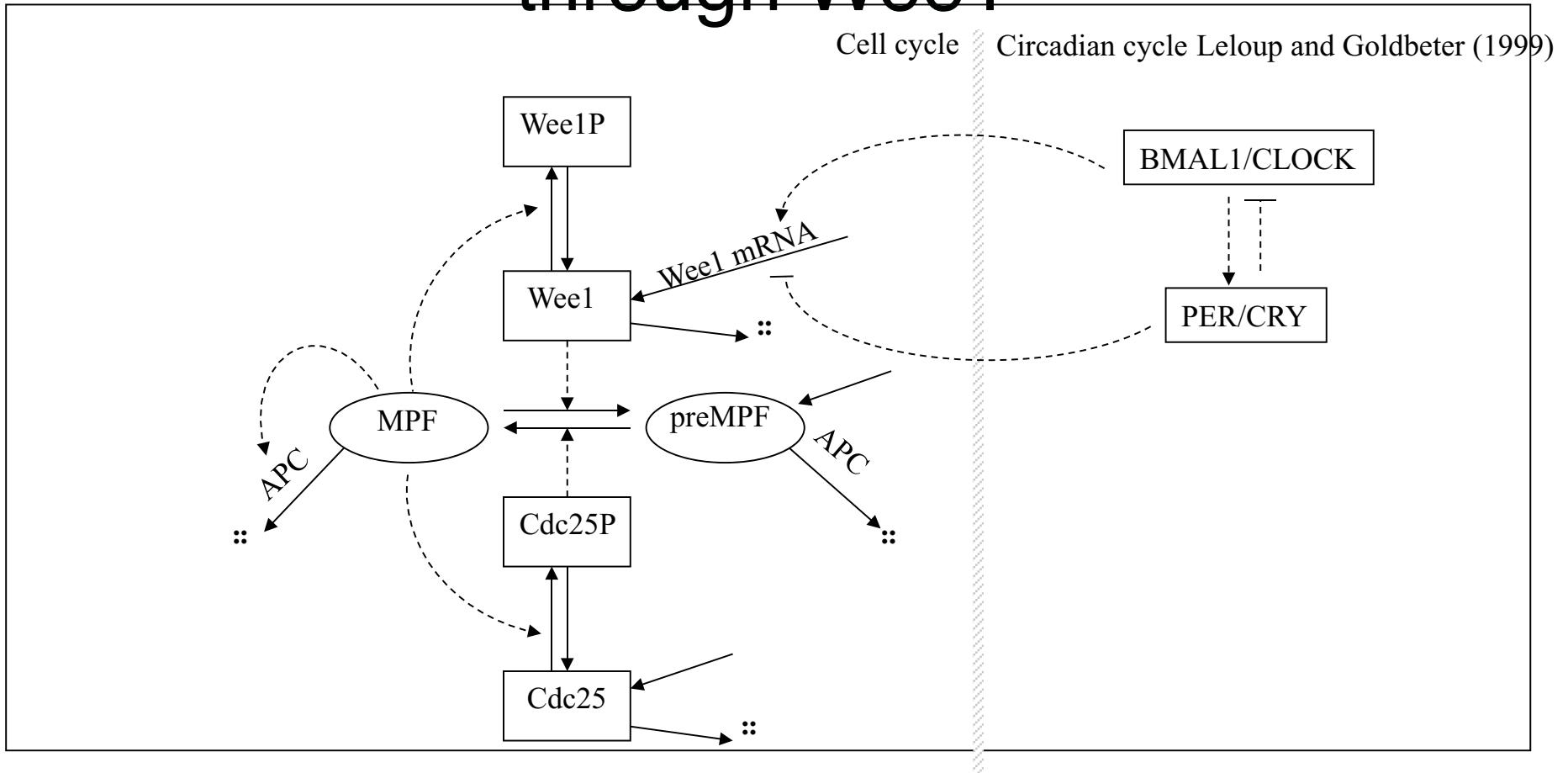
Parameter Search from Temporal Properties

```
biocham: learn_parameter([k3,k4],[(0,200),(0,200)],20,  
period(Cdc2-Cyclin~{p1},35), 150).
```

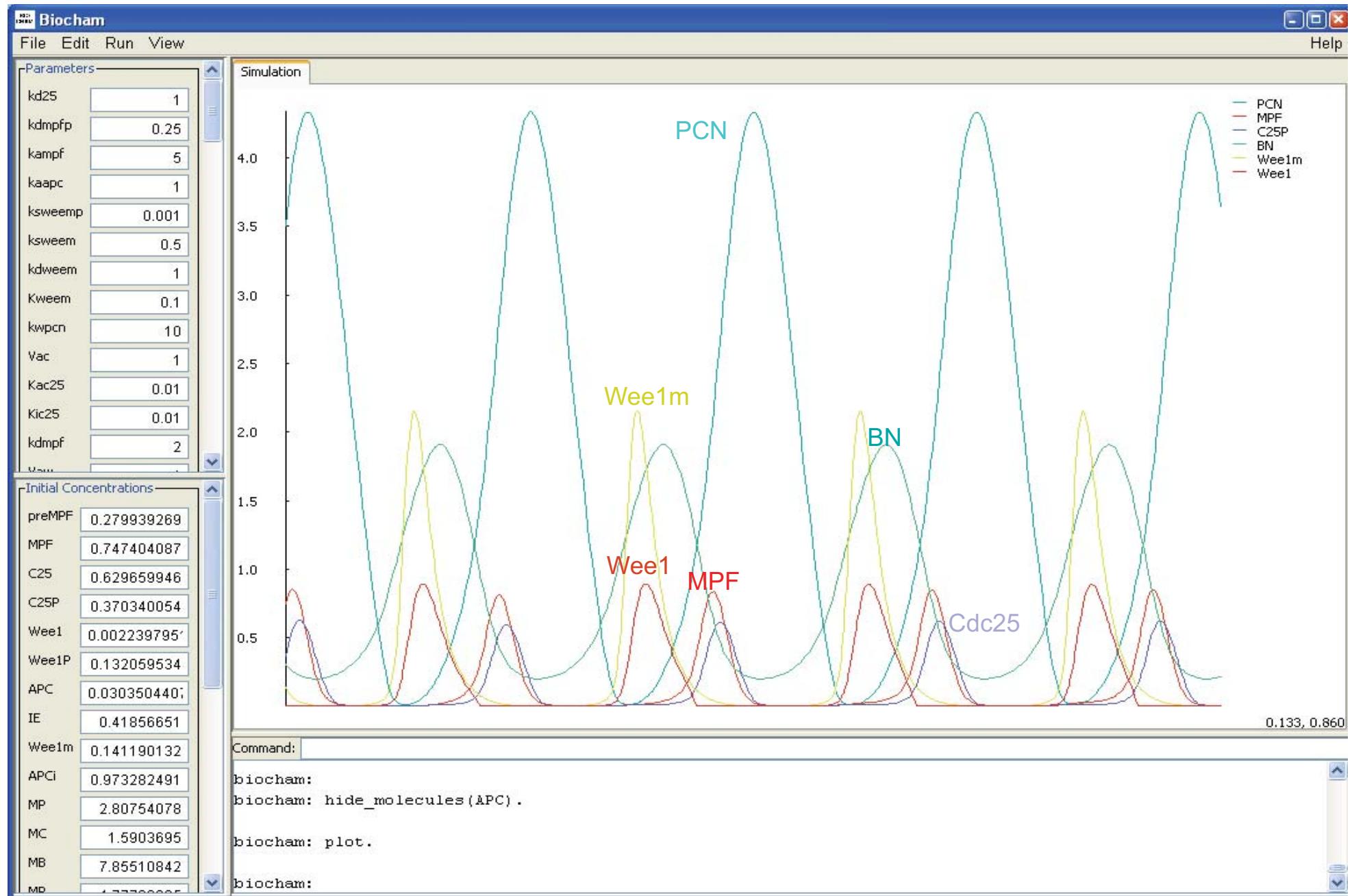
First values found:
parameter(k3,10).
parameter(k4,280).



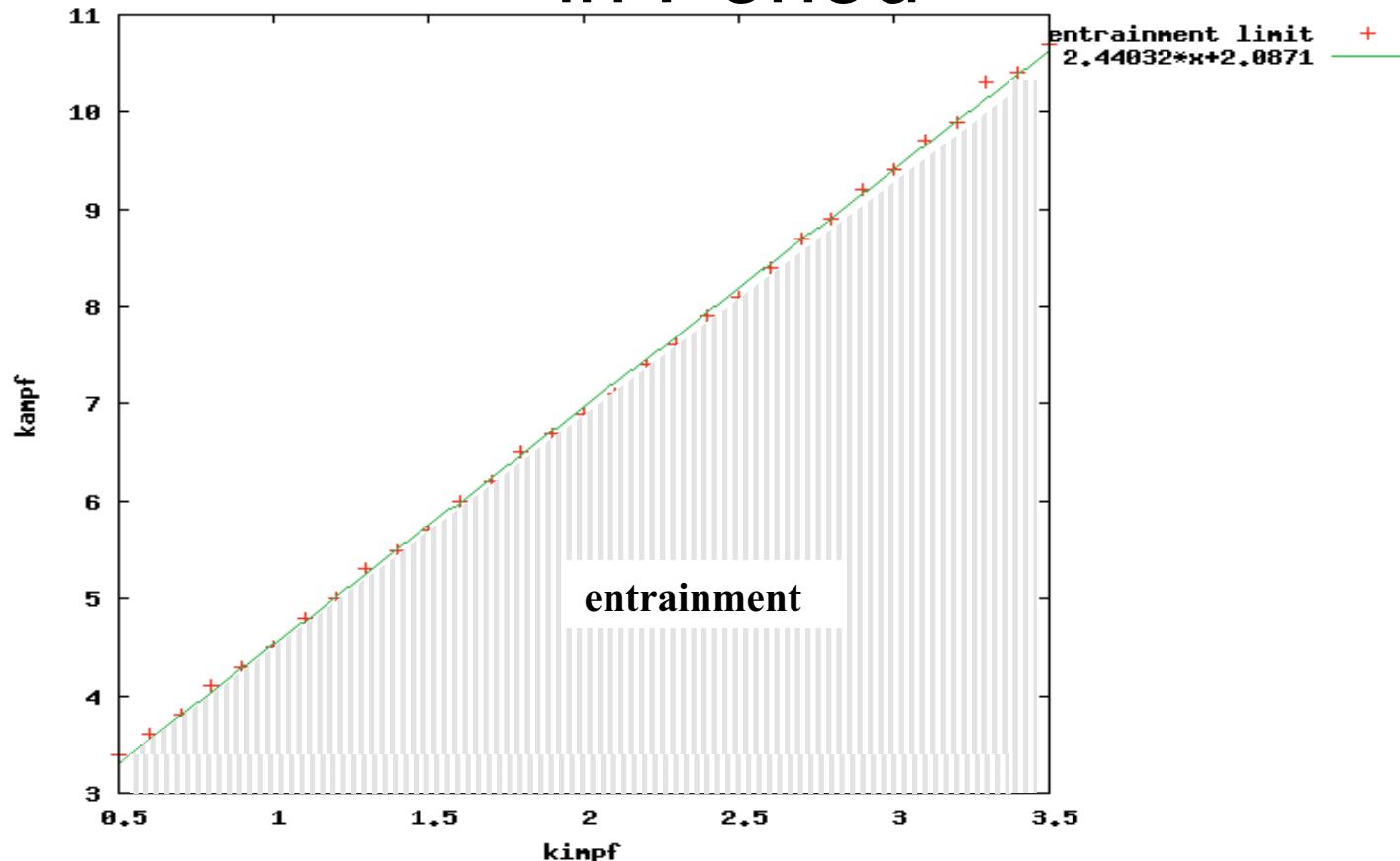
Linking the Cell and Circadian Cycles through Wee1



[L. Calzone, S. Soliman 2006]



Condition on Wee1/Cdc25 for the Entrainment in Period



Entrainment in period constraint expressed in LTL with the period formula

True/False valuation of temporal logic formulae

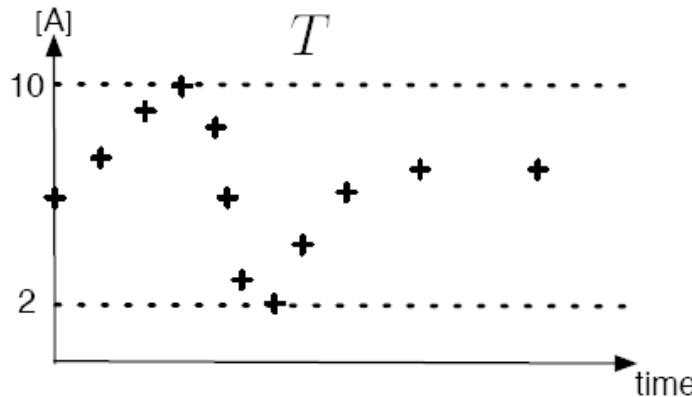
The **True/False** valuation of temporal logic formulae is **not well adapted** to several problems :

- parameter search, optimization and control of continuous models
- quantitative estimation of robustness
- sensitivity analyses

→ need for a continuous degree of satisfaction of temporal logic formulae

How far is the system from verifying the specification ?

Model-Checking Generalized to Constraint Solving



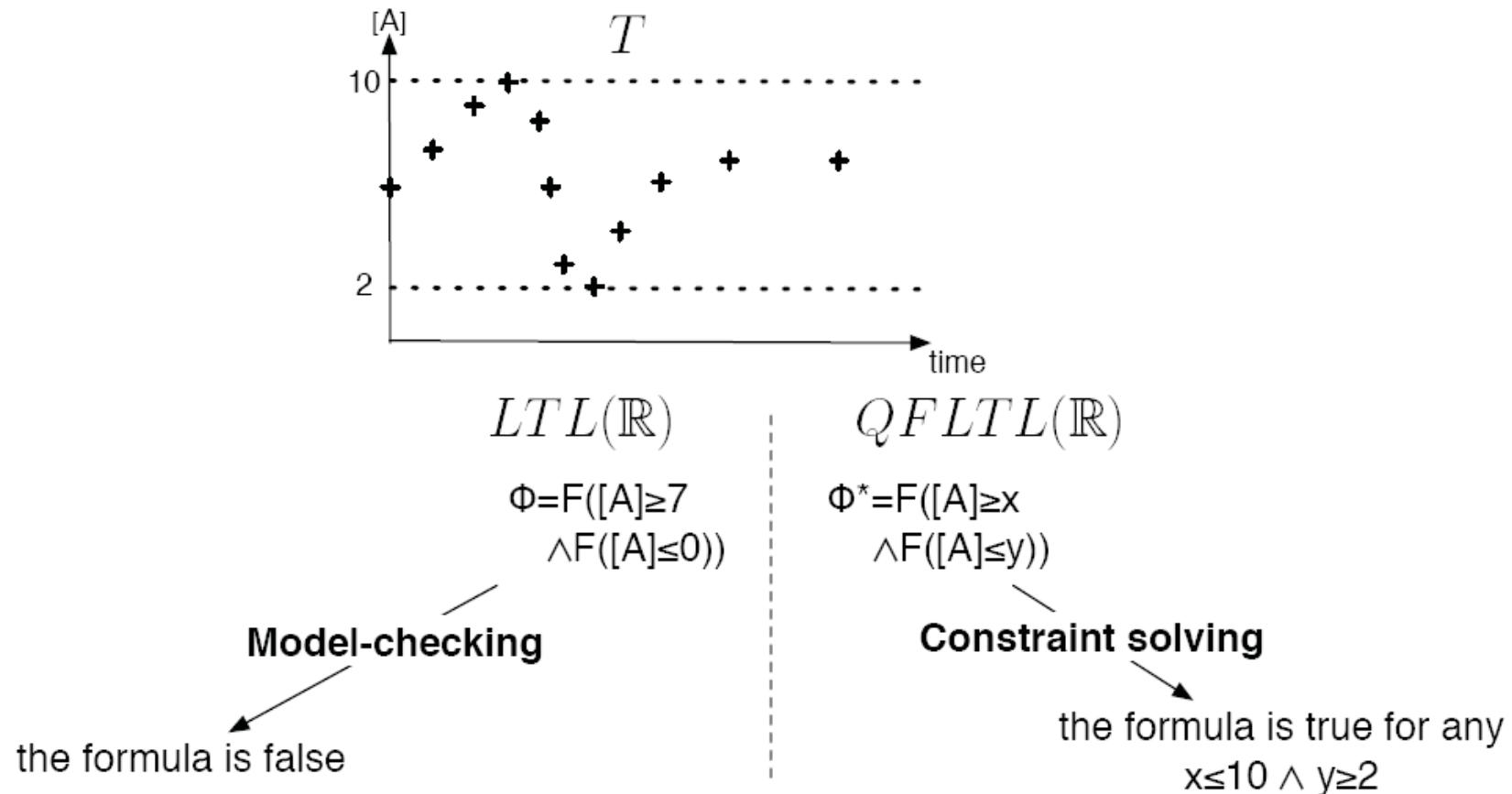
$LTL(\mathbb{R})$

$$\Phi = F([A] \geq 7 \wedge F([A] \leq 0))$$

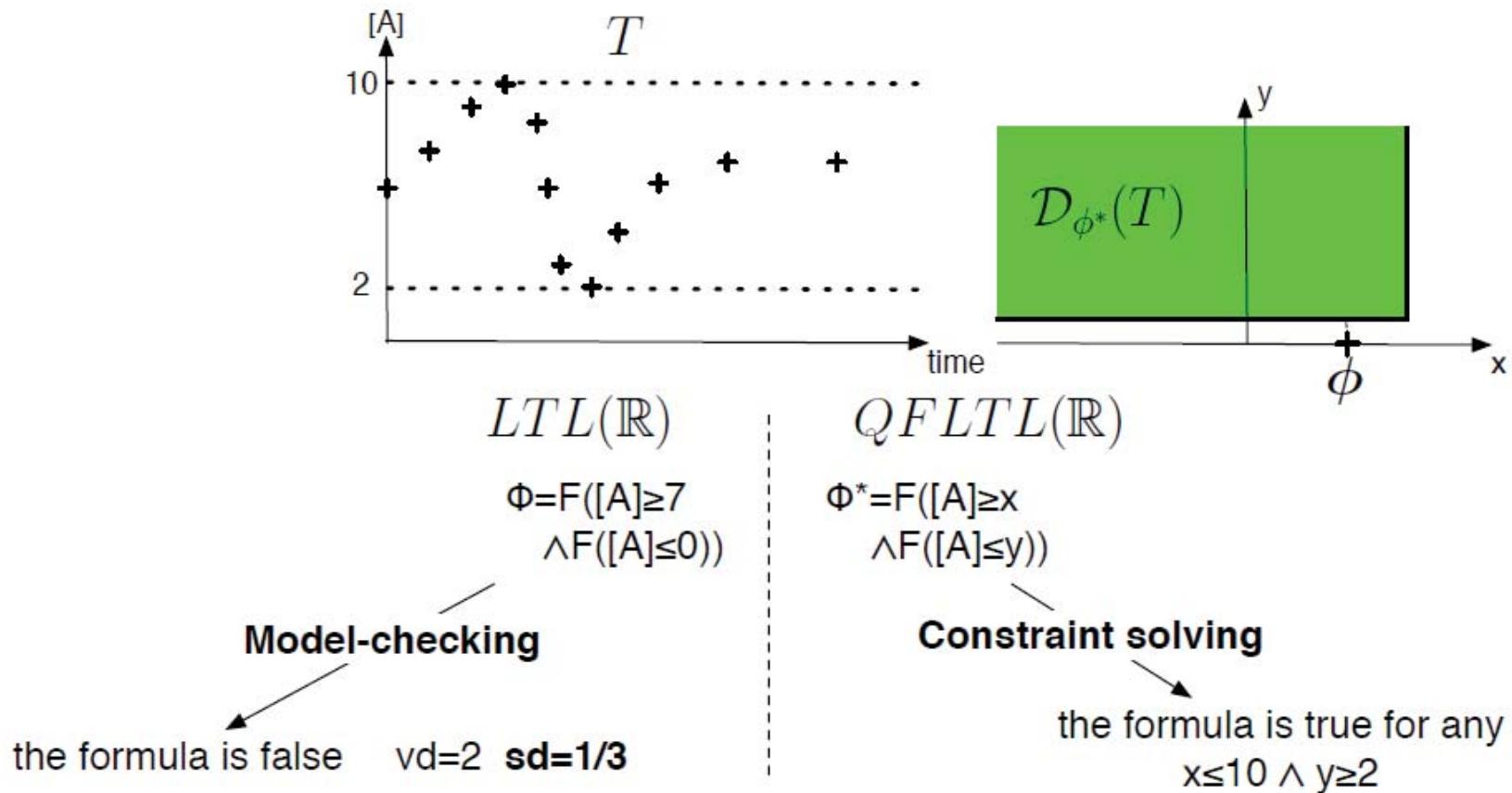
Model-checking

the formula is false

Model-Checking Generalized to Constraint Solving



Model-Checking Generalized to Constraint Solving



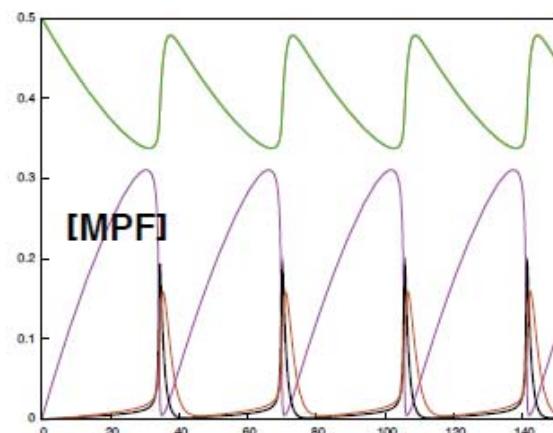
Validity domain $\mathcal{D}_{\phi^*}(T)$ for the **free variables** in ϕ^* [Fages Rizk CMSB'07]

Violation degree $vd(T, \phi) = \text{distance}(\text{val}(\phi), \mathcal{D}_{\phi^*}(T))$

Satisfaction degree $sd(T, \phi) = \frac{1}{1+vd(T, \phi)} \in [0, 1]$

Learning kinetic parameter values from LTL specifications

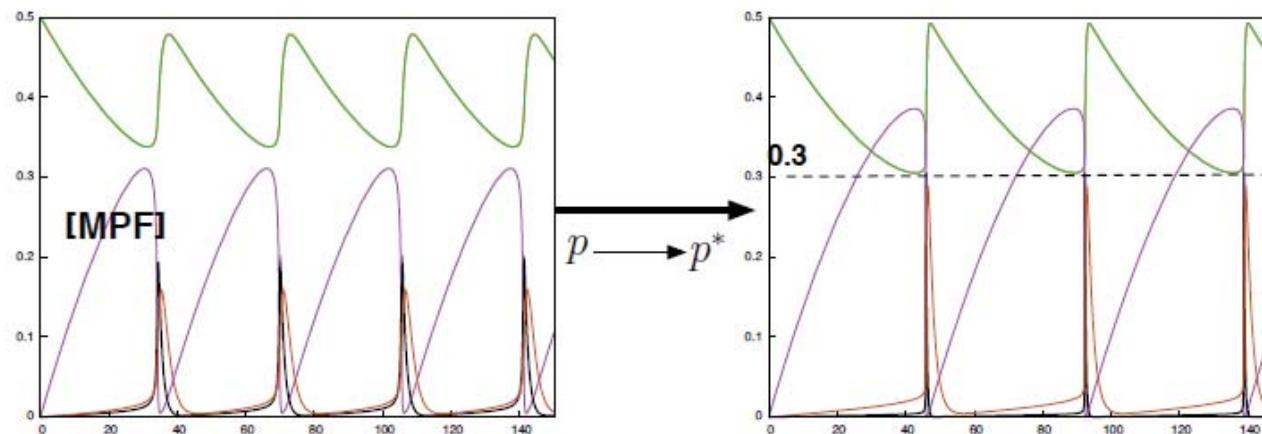
- simple model of the yeast cell cycle from [Tyson PNAS 91]
- models Cdc2 and Cyclin interactions (6 variables, 8 kinetic parameters)



- Pb : find values of 8 parameters such that amplitude is ≥ 0.3
 $\phi^*: \mathbf{F}([A]>x \wedge \mathbf{F}([A]<y))$
amplitude $z=x-y$
goal : $z = 0.3$

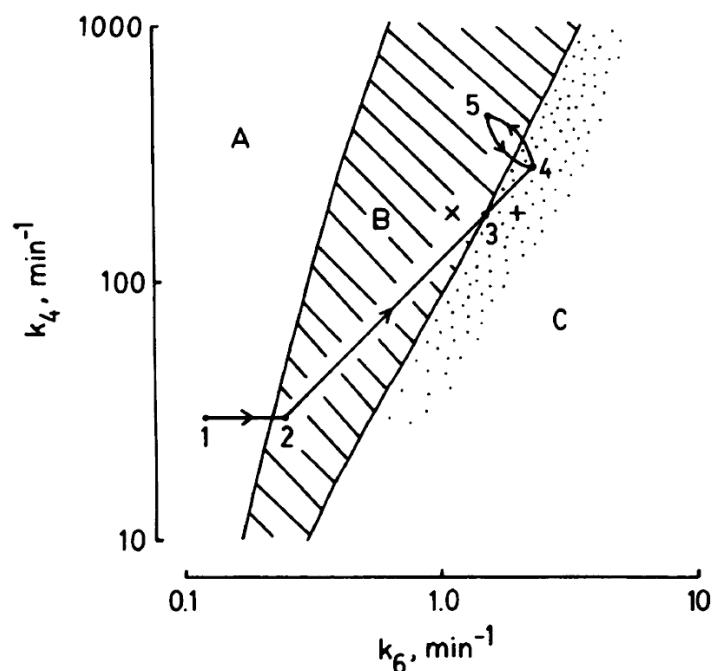
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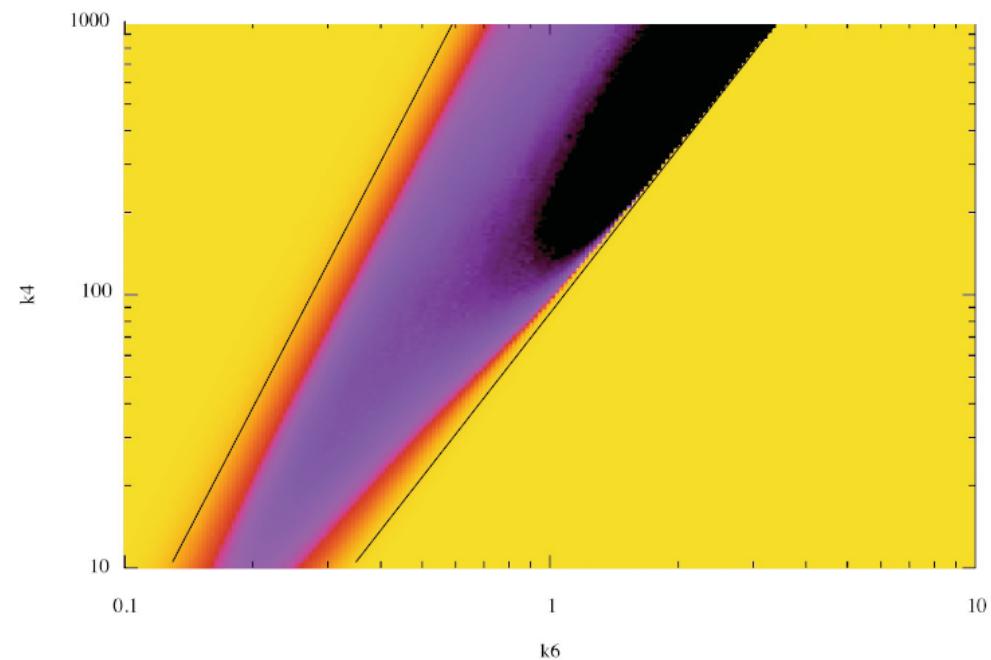


- Pb : find values of 8 parameters such that amplitude is ≥ 0.3
 $\phi^* : \mathbf{F}([A]>x \wedge \mathbf{F}([A]<y))$
amplitude $z=x-y$
goal : $z = 0.3$
- → solution found after 30s (100 calls to the fitness function)

LTL(R) Satisfaction Degree and Bifurcation Diagram



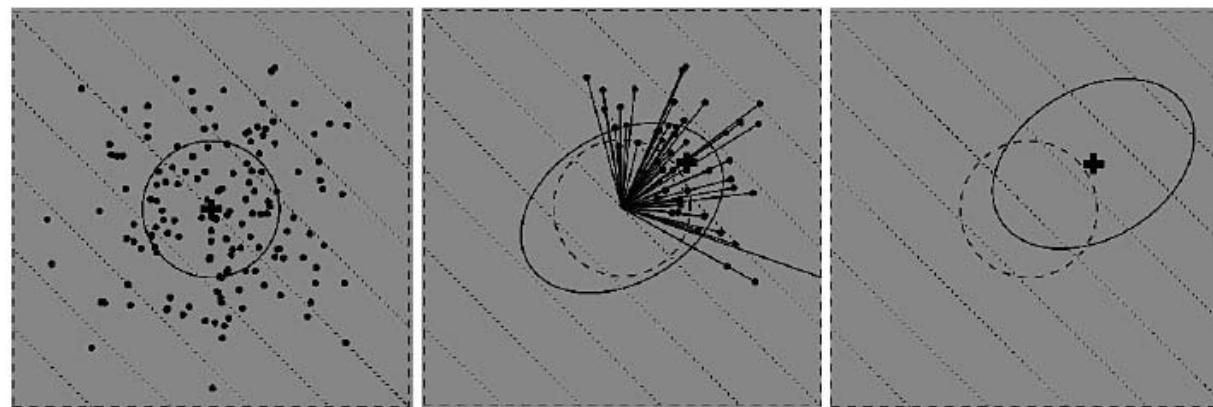
Bifurcation diagram on k_4, k_6
[Tyson 91]



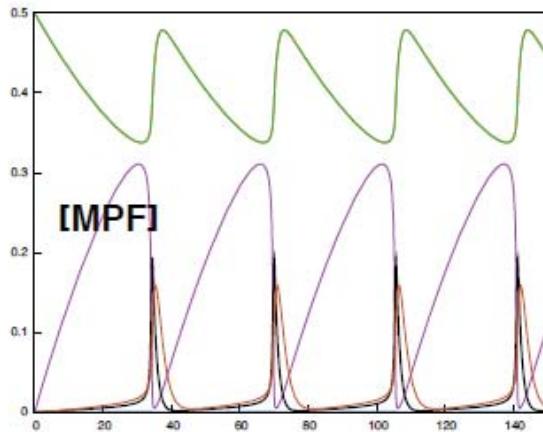
Continuous satisfaction degree in $[0,1]$
of an LTL(R) formula for oscillation
with amplitude constraint
[Rizk Batt Fages Soliman CMSB 08]

Black-box Randomized Non-linear Optimization Method

- Use existing non-linear optimization toolbox for kinetic parameter search using satisfaction degree as fitness function
- We use the state-of-the-art Covariance Matrix Adaptation Evolution Strategy (CMA-ES) [Hansen Osermeier 01, Hansen 08]
- CMA-ES maximizes an objective function in continuous domain in a black box scenario
- CMA-ES uses a probabilistic neighborhood and updates information in covariance matrix at each move

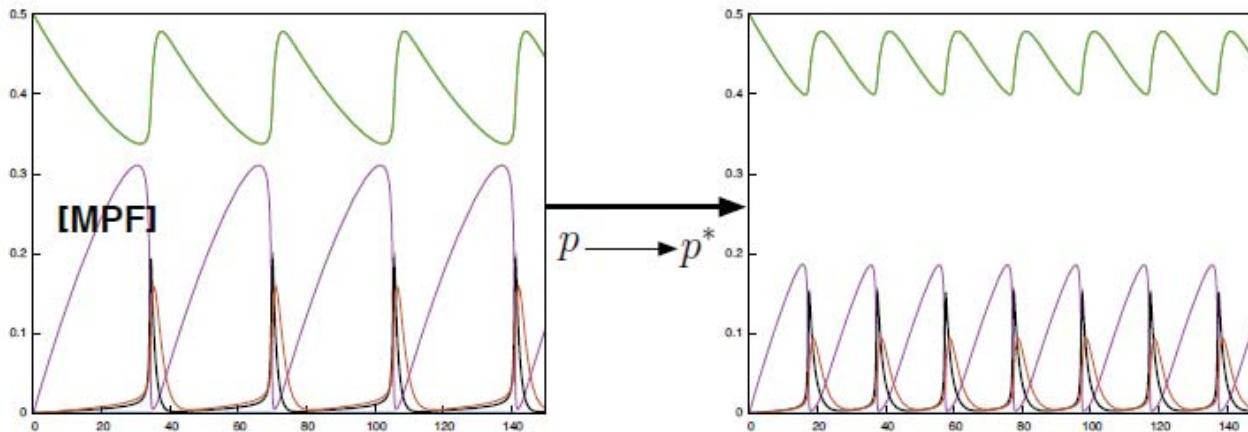


Learning Parameter Values from Period Constraints in LTL



- Pb : find values of 8 parameters such that period is 20
 $\phi^* : \mathbf{F}(\text{MPF}_{localmaximum} \wedge \text{Time}=t1 \wedge \mathbf{F}(\text{MPF}_{localmaximum} \wedge \text{Time}=t2))$
(with $\text{MPF}_{localmaximum}$: $d([\text{MPF}])/dt > 0 \wedge \mathbf{X}(d([\text{MPF}])/dt < 0)$)
period $z=t2-t1$
goal $z=20$

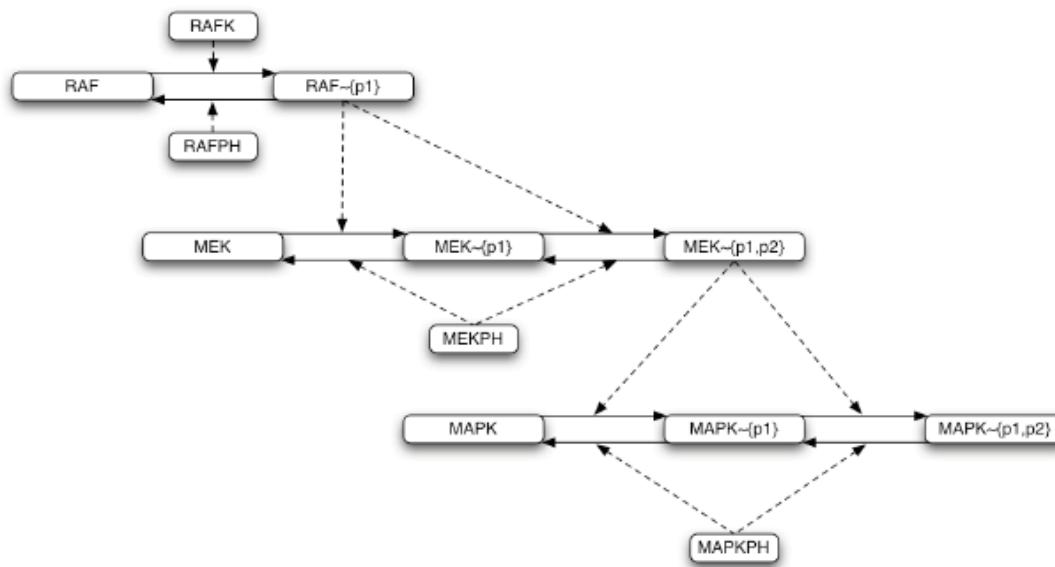
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(with $\text{MPF}_{localmaximum}$: $d([\text{MPF}])/dt > 0 \wedge \mathbf{X}(d([\text{MPF}])/dt < 0)$)
period $z=t2-t1$
goal $z=20$
- → Solution found after 60s (200 calls to the fitness function)

Oscillations in MAPK signal transduction cascade

- MAPK signaling model [Huang Ferrel PNAS 93]



- **search for oscillations in 37 dimensions** (30 parameters and 7 initial conditions)
→ solution found after 3 min (200 calls to the fitness function)
Oscillations already observed by simulation [Qiao et al. 07]
- No negative feedback in the **reaction graph**, but negative circuits in the **influence graph** [Fages Soliman FMSB'08, CMSB'06]

Robustness Measure Definition

Robustness defined with respect to :

- a biological system
- a functionality property D_a
- a set P of perturbations
- General notion of robustness proposed in [Kitano MSB 07]:

$$\mathcal{R}_{a,P} = \int_{p \in P} D_a(p) \ prob(p) \ dp$$

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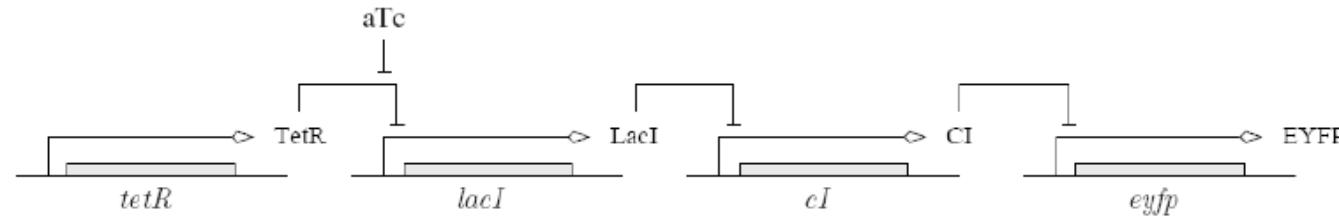
- Our computational measure of robustness w.r.t. LTL(\mathbb{R}) spec:
Given an ODE model with initial conditions, a TL formula ϕ and a set of perturbations P (on initial conditions or parameters),

$$\mathcal{R}_{\phi,P} = \int_{p \in P} sd(T(p), \phi) \ prob(p) \ dp$$

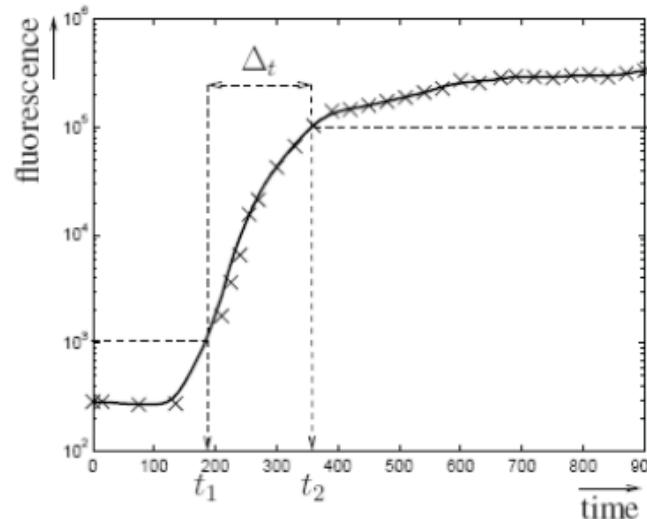
where $T(p)$ is the trace obtained by numerical integration of the ODE for perturbation p

Application to Synthetic Biology in *E. Coli*

Cascade of transcriptional inhibitions implemented in *E.coli* [Weiss et al PNAS 05]



The output protein EYFP is controlled by the small input molecule aTc



The system is well-timed if EYFP remains below 10^3 for at least 150 min., then exceeds 10^5 after at most 450 min., and switches from low to high levels in less than 150 min.

Specifying the expected behavior in LTL(\mathbb{R})

The timing specifications can be formalized in temporal logic as follows:

$$\begin{aligned}\phi(t_1, t_2) = & \quad \mathbf{G}(\text{time} < t_1 \rightarrow [\text{EYFP}] < 10^3) \\ & \wedge \mathbf{G}(\text{time} > t_2 \rightarrow [\text{EYFP}] > 10^5) \\ & \wedge t_1 > 150 \wedge t_2 < 450 \wedge t_2 - t_1 < 150\end{aligned}$$

which is abstracted into

$$\begin{aligned}\phi(t_1, t_2, b_1, b_2, b_3) = & \quad \mathbf{G}(\text{time} < t_1 \rightarrow [\text{EYFP}] < 10^3) \\ & \wedge \mathbf{G}(\text{time} > t_2 \rightarrow [\text{EYFP}] > 10^5) \\ & \wedge t_1 > b_1 \wedge t_2 < b_2 \wedge t_2 - t_1 < b_3\end{aligned}$$

for computing validity domains for b_1, b_2, b_3

with the objective $b_1 = 150, b_2 = 450, b_3 = 150$ for computing the satisfaction degree in a given trace.

Improving robustness

Variance-based global sensitivity indices

$$S_i = \frac{\text{Var}(E(R|P_i))}{\text{Var}(R)} \in [0, 1]$$

S_γ	20.2 %	$S_{\kappa_{eyfp}, \gamma}$	8.7 %
$S_{\kappa_{eyfp}}$	7.4 %	$S_{\kappa_{cl}, \gamma}$	6.2 %
$S_{\kappa_{cl}}$	6.1 %	$S_{\kappa_{cl}^0, \gamma}$	5.0 %
$S_{\kappa_{lacI}^0}$	3.3 %	$S_{\kappa_{cl}^0, \kappa_{eyfp}}$	2.8 %
$S_{\kappa_{cl}^0}$	2.0 %	$S_{\kappa_{cl}, \kappa_{eyfp}}$	1.8 %
$S_{\kappa_{lacI}}$	1.5 %	$S_{\kappa_{eyfp}^0, \gamma}$	1.5 %
$S_{\kappa_{eyfp}^0}$	0.9 %	$S_{\kappa_{cl}^0, \kappa_{cl}}$	1.1 %
$S_{u_{aTc}}$	0.4 %	$S_{\kappa_{cl}^0, \kappa_{lacI}}$	0.5 %
total first order	40.7 %	total second order	31.2 %

degradation factor γ has the strongest impact on the cascade.

aTc variations have a very low impact

different importance of the basal κ_{eyfp}^0 and regulated κ_{eyfp} EYFP production rates

the basal production of EYFP is due to an incomplete repression of the promoter by CI (high effect of κ_{cl}) rather than a constitutive leakage of the promoter (low effect of κ_{eyfp}^0). [Rizk Batt Fages Soliman ISMB'09 Bioinformatics]

Conclusion

- New focus in Systems Biology: formal methods from Computer Science
 - Beyond diagrammatic notations: formal semantics, abstract interpretation
 - Beyond curve fitting: high-level specifications in temporal logic
 - Automatic model-checking. Parameter optimization. Model reduction

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 - Beyond discrete machines: stochastic or continuous or hybrid dynamics
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- New focus in Programming: numerical methods
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 - Quantitative transition systems
 - Temporal logic constraint solving, continuous satisfaction degree, optimization
- Synthetic Biology
 - Program the living with programming tools
 - Computational design of gene circuits

Acknowledgments

- **Contraintes group** at INRIA Paris-Rocquencourt on this topic:
Grégory Batt, Sylvain Soliman, Elisabetta De Maria, Aurélien Rizk, Steven Gay, Faten Nabli, Xavier Duportet, Janis Ulhendorf, Dragana Jovanovska
- **EraNet SysBio C5Sys** (follow up of FP6 Tempo) on cancer chronotherapies, coord.
Francis Lévi, INSERM; Jean Clairambault INRIA; ...
Coupled models of cell and circadian cycles, p53/mdm2, cytotoxic drugs.
- **INRIA/INRA project Regate** coord. *F. Clément INRIA; E. Reiter, D. Heitzler*
Modeling of GPCR Angiotensine and FSH signaling networks
- **ANR project Calamar**, coord. *C. Chaouiya, D. Thieffry , ENS, L. Calzone, Curie...*
Modularity and Compositionality in regulatory networks.
- **OSEO Biointelligence**, coord. *Dassault-Systèmes,*
Technology transfer of Biocham concepts and tools

T7 Bacteriophage Infection Process

Retrovirus:

- Template nucleic acids (RNA): tem (initial condition of low infection)
- Genomic nucleic acids (DNA): gen
- Structural proteins: struc

MA(c1) for gen => tem.

parameter(c1,0.025).

MA(c2) for tem => _.

parameter(c2,0.25).

MA(c3) for tem => tem+gen.

parameter(c3,1.0).

MA(c4) for gen+struc => virus.

parameter(c4,0.0000075).

MA(c5) for tem => tem+struc.

parameter(c5,1000).

MA(c6) for struc => _.

parameter(c6,1.99).

T7 Bacteriophage ODE semantics

MA(c1) for gen => tem.

MA(c2) for tem => _.

MA(c3) for tem => tem+gen.

MA(c4) for gen+struc => virus.

MA(c5) for tem => tem+struc.

MA(c6) for struc => _.

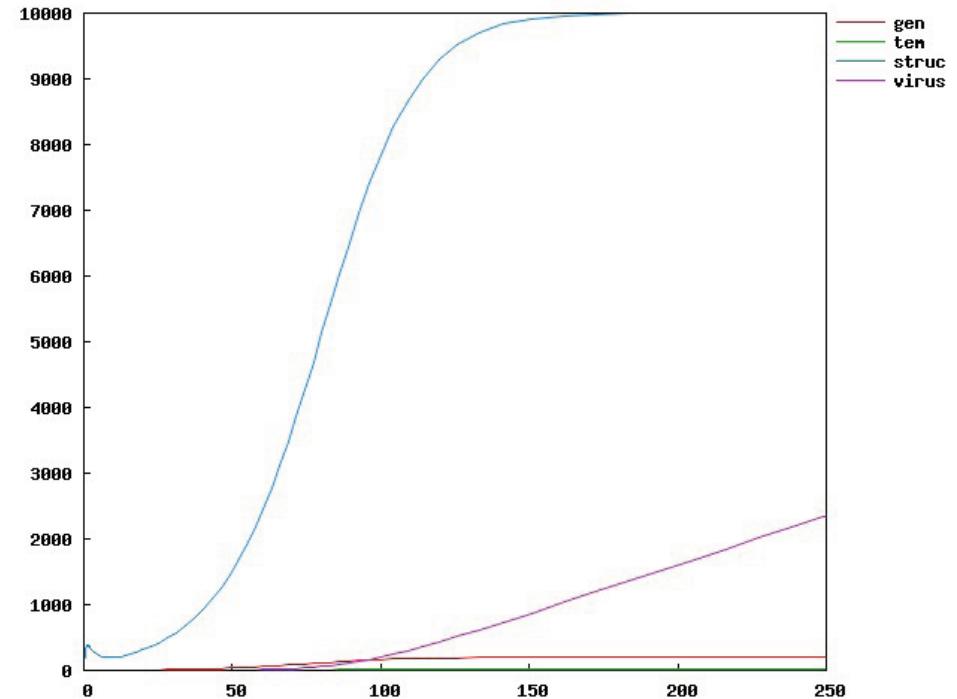
[tem] = 1

$d[virus]/dt = c4 * [gen] * [struc]$

$d[tem]/dt = c1 * [gen] - c2 * [tem]$

$d[struc]/dt = c5 * [tem] - c6 * [struc] - c4 * [gen] * [struc]$

$d[gen]/dt = c3 * [tem] - c4 * [gen] * [struc] - c1 * [gen]$



T7 Bacteriophage ODE semantics

2 steady states $dX/dt=0$:

1) tem=gen=struc=0

2) tem=20 gen=200 struc=10000

cyclic behavior

Minimal T-invariants in Petri nets

$I^*V=0$ (incidence stoichiometric matrix)

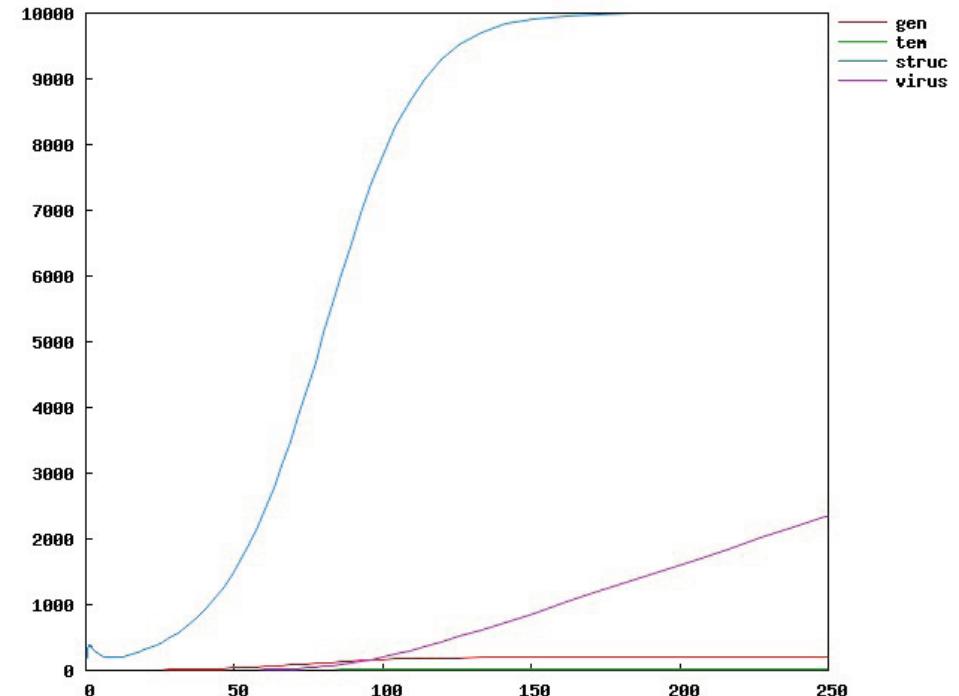
Elementary Modes of flux distribution

in metabolic networks

$$\text{tem} = - (c_1 * c_6) / (- (c_2 * c_5) - c_1 * c_5 + c_3 * c_5) * (c_2 * c_4 / (- (c_1 * c_3) + c_2 * c_3))^{-1}$$

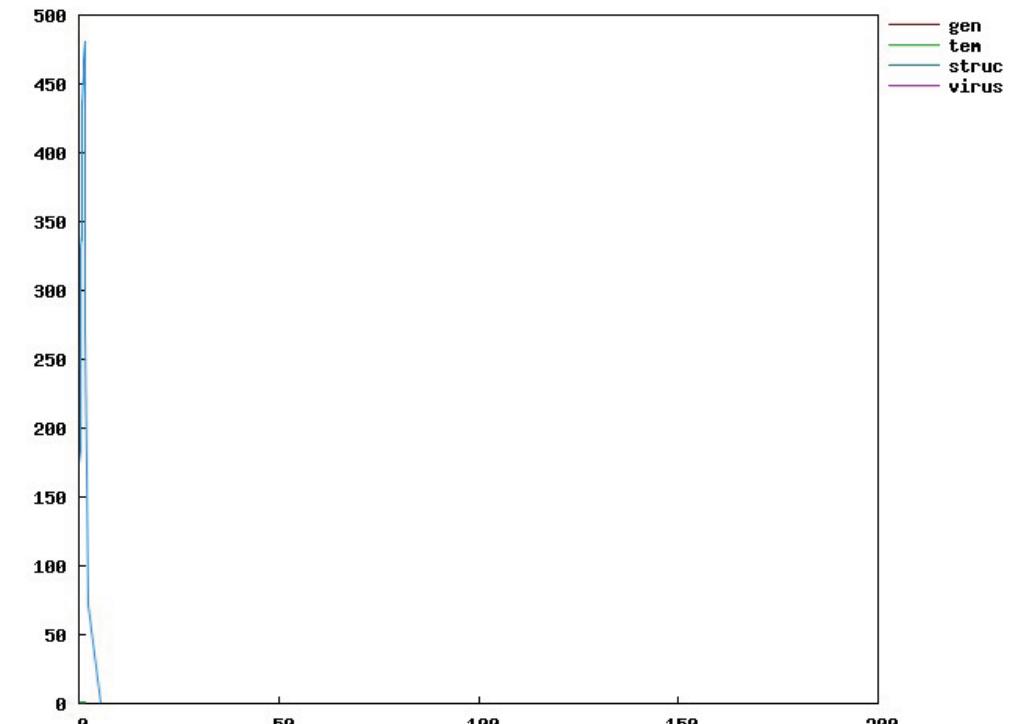
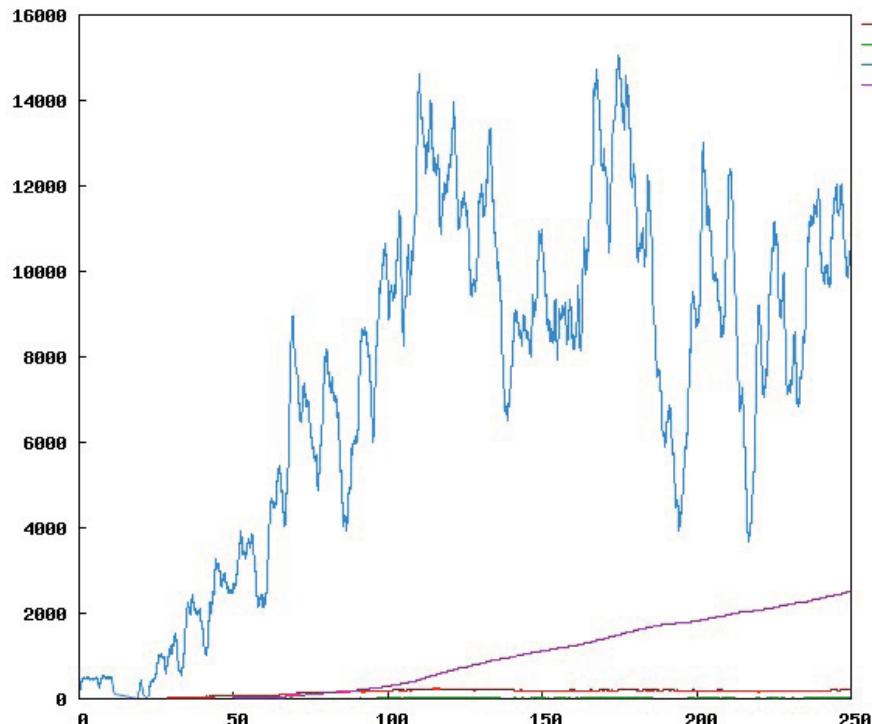
$$\text{gen} = ((c_2 * c_6 / (- (c_3 * c_5) + c_2 * c_5 + c_1 * c_5))^{-1} * (c_2 * c_4 / (- (c_1 * c_3) + c_2 * c_3)))^{-1}$$

$$\text{struc} = (c_2 * c_4 / (- (c_1 * c_3) + c_2 * c_3))^{-1}$$



T7 Bacteriophage Stochastic semantics

Two qualitatively different behaviors:



T7 Bacteriophage Boolean Semantics

Querying all possible behaviors in Computation Tree Logic
(symbolic model-checking NuSMV)

biocham: nusmv(EF (!(gen) & !(tem) & !(struc) & !(virus))).

true

biocham: why.

tem is present

2 tem => _.

tem is absent

Query time: 0.00 s

T7 Bacteriophage Boolean Semantics

biocham: nusmv(EF (virus & EF (!(gen) & !(tem) & !(struc)))).

true

biocham: why.

tem is present

5 *tem => struc+tem.*

struc is present

3 *tem => gen+tem.*

gen is present

4 *gen+struc => virus.*

gen is absent

struc is absent

virus is present

2 *tem => _.*

tem is absent

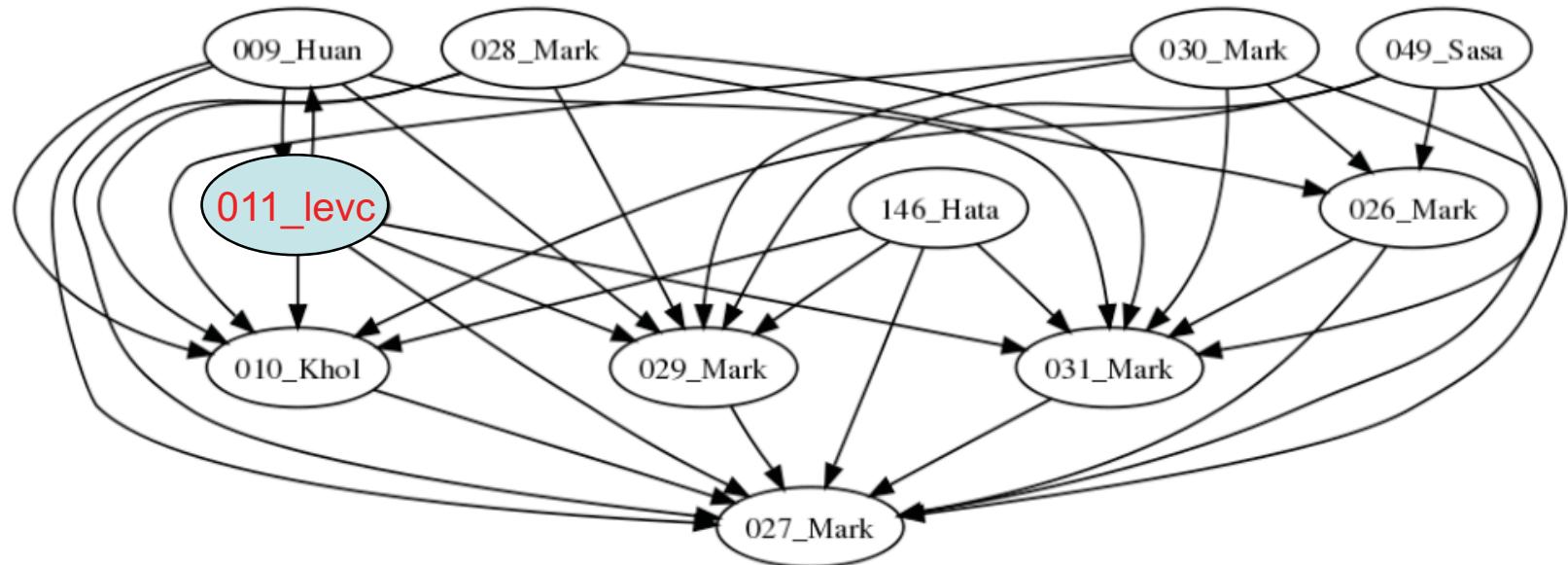
Aut. Generation of Temporal Logic Properties

Ei(reachable(gen))
Ei(reachable(!(gen)))
Ei(steady(!(gen)))
Ai(checkpoint(tem,gen)))
Ei(reachable(tem))
Ei(reachable(!(tem)))
Ei(steady(tem))
Ai(checkpoint(gen,tem)))
Ei(reachable(struc))
Ei(reachable(!(struc)))
Ei(steady(!(struc)))

Ai(checkpoint(tem,struc)))
Ei(reachable(virus))
Ei(reachable(!(virus)))
Ei(steady(!(virus)))
Ai(checkpoint(gen,virus)))
Ai(checkpoint(gen,!(virus))))
Ai(checkpoint(tem,!(virus))))
Ai(checkpoint(struc,virus)))

Time: 0.06 s

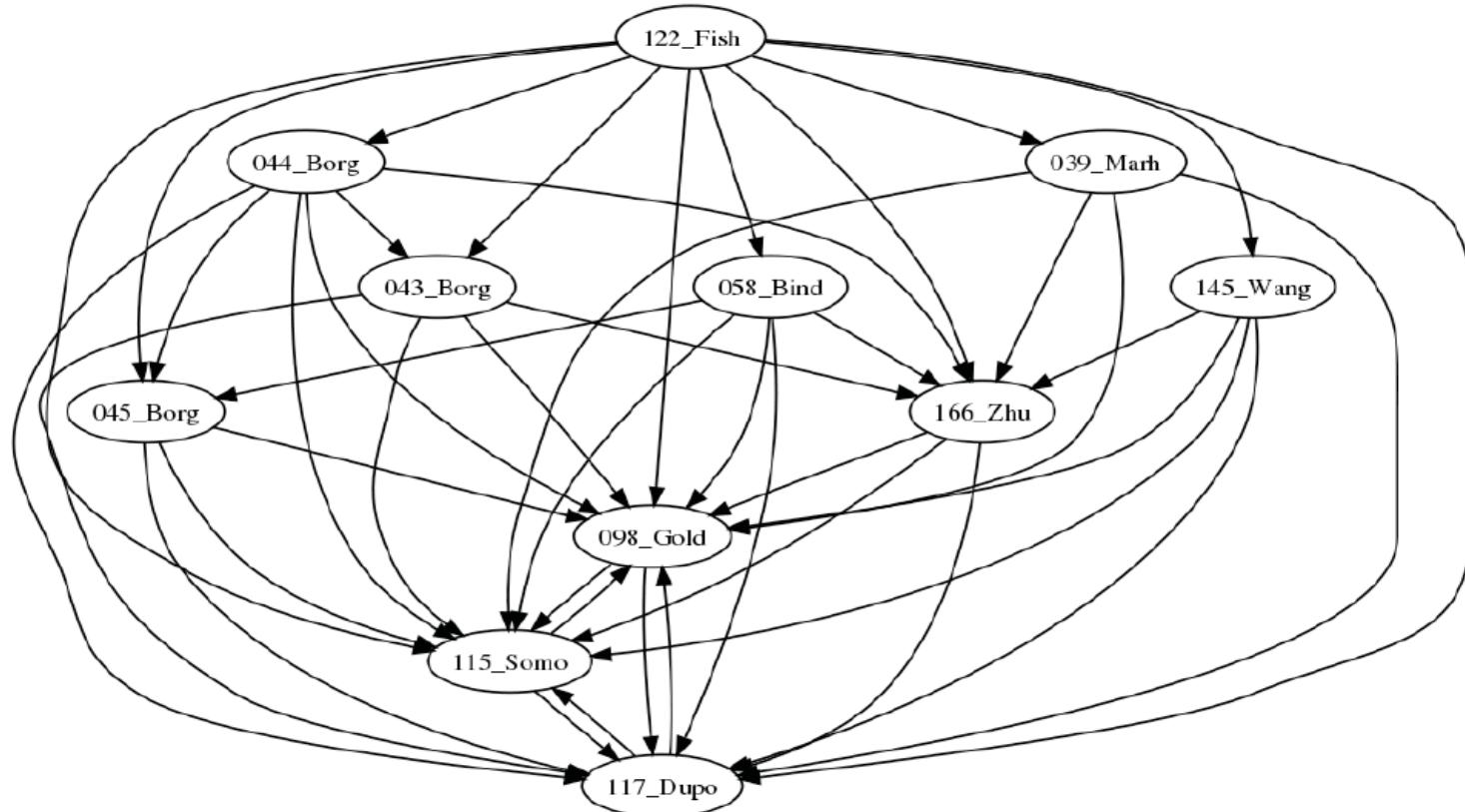
Hierarchies of Models



MAPK models from SBML model repository <http://www.biomodels.net>
[Gay Fages Soliman, A graph matching method for reducing and relating models 2010 biionfo]

- 4 graph operations: delete/merge species/reactions
- Model reductions/refinements as subgraph epimorphisms
- Query language for model repositories (constraint program)

Hierarchies of Models

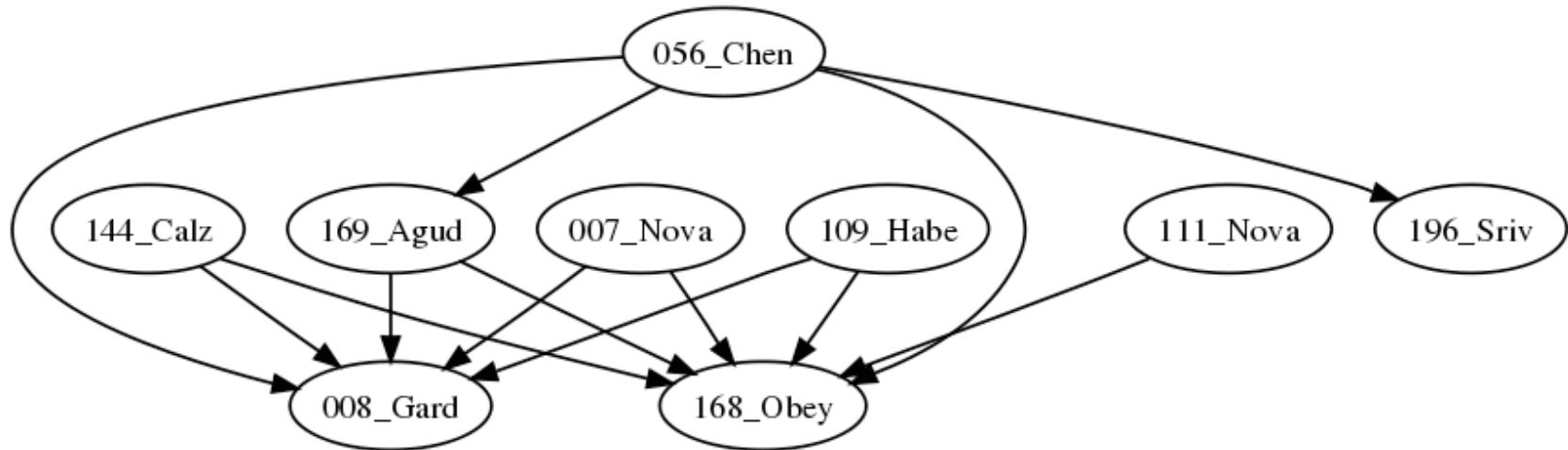


Models of circadian clock in <http://www.biomodels.net>

Computation of subgraph epimorphisms by constraint programming

Scales up to most of the 241^2 pairs of models in biomodels.

Hierarchies of Models



- Cell cycle models from biomodels.net
- ODE models not well transcribed in SBML (missing reactants)
- Imperfect model reductions found by graph matching

A Programmer View at Cell Computations

Size of genome

- 5 Mb for bacteria: normal size program (Biocham binary: 15Mb as yeast)
- 3 Gb for human: normal size of a video not for a program
- 140 Gb for lung fish: nature error !

Speed of interactions

- Protein interactions: enzyme-substrate collisions at 0,5 Mhz, quite slow
- Gene expression: hours ! as slow as reinstalling an operating system

Concurrent computation paradigm

- Chemical metaphor for concurrent programming [Banatre, Le Metayer 86]
- CHAM [Berry Boudol 90] to express the operational semantics of the Pi-Calculus
- Membranes for modules: just like cell compartments

Hybrid continuous+discrete computations (*energy + information*)

- Trend for future: more *physics* in *informatics*, more *informatics* in *physics*